

UDC 616-056.527+618.3-06+159.922.1-055.26+577.121+618.36-008.64
DOI: 10.56871/CmN-W.2023.40.28.001

THE PREDICTIVE ROLE OF OBESITY FOR MATERNAL AND CHILD HEALTH IN THE FIRST YEAR OF LIFE

© Dmitry O. Ivanov, Natalya E. Prokopeva, Yury V. Petrenko

Saint Petersburg State Pediatric Medical University. Lithuania 2, Saint Petersburg, Russian Federation, 194100

Contact information:

Natalya E. Prokopeva — Laboratory assistant-researcher of the laboratory Medical and Social problems in pediatrics, Research Center of Saint Petersburg State Pediatric Medical University. E-mail: shkumat93@gmail.com ORCID ID: 0000-0001-5412-1412

For citation: Ivanov DO, Prokopeva NE, Petrenko YuV. The predictive role of obesity for maternal and child health in the first year of life. Children's medicine of the North-West (St. Petersburg). 2023; 11(3): 5-35. DOI: <https://doi.org/10.56871/CmN-W.2023.40.28.001>

Received: 01.06.2023

Revised: 03.08.2023

Accepted: 12.09.2023

Abstract. Over the past few decades, the prevalence of obesity worldwide has reached epidemic proportions. Obesity and overweight during pregnancy are associated with worse maternal and child outcomes. In addition, studies show that maternal obesity can lead to long-term consequences for the offspring, increasing the risk of neuropsychiatric disorders, metabolic, atopic diseases, and possible changes in the immune / inflammatory status. In addition to genetic mechanisms, a growing body of evidence suggests the induction of epigenetic changes by maternal obesity, which may influence offspring phenotype, thereby influencing later risk of obesity and cardiometabolic disease. However, the mechanisms linking the maternal environment to adverse short and long term outcomes remain poorly understood. This review presents current knowledge about the impact of maternal obesity on a child in the first year of life. Understanding these processes is key to developing therapeutic interventions to prevent future cardiovascular and metabolic pathologies in future generations.

Key words: maternal obesity; perinatal period; metabolic syndrome; offspring of obese mothers; mother-placenta-fetus.

ПРОГНОСТИЧЕСКАЯ РОЛЬ ОЖИРЕНИЯ ДЛЯ ЗДОРОВЬЯ МАТЕРИ И РЕБЕНКА В ПЕРВЫЙ ГОД ЖИЗНИ

© Дмитрий Олегович Иванов, Наталья Эдуардовна Прокопьева, Юрий Валентинович Петренко

Санкт-Петербургский государственный педиатрический медицинский университет. 194100, г. Санкт-Петербург, ул. Литовская, 2

Контактная информация:

Наталья Эдуардовна Прокопьева — лаборант-исследователь лаборатории Медико-социальных проблем в педиатрии, НИЦ ФГБОУ ВО СПбГПМУ. E-mail: shkumat93@gmail.com ORCID ID: 0000-0001-5412-1412

Для цитирования: Иванов Д.О., Прокопьева Н.Э., Петренко Ю.В. Прогностическая роль ожирения для здоровья матери и ребенка в первый год жизни // Children's medicine of the North-West. 2023. Т. 11. № 3. С. 5–35. DOI: <https://doi.org/10.56871/CmN-W.2023.40.28.001>

Поступила: 01.06.2023

Одобрена: 03.08.2023

Принята к печати: 12.09.2023

Резюме. За последние несколько десятилетий распространенность ожирения во всем мире достигла масштабов эпидемии. Ожирение и избыточный вес во время беременности связаны с ухудшением исходов для матери и ребенка. Кроме того, исследования показывают, что материнское ожирение может привести к долгосрочным последствиям для потомства, увеличивая риск развития психоневрологических расстройств, метаболических, атопических заболеваний, а также возможных изменений иммунного / воспалительного статуса. В дополнение к генетическим механизмам все больше данных свидетельствует об индукции эпигенетических изменений материнским ожирением, которые могут влиять на фенотип потомства, тем самым программируя риск ожирения и кардиометаболических заболеваний. Однако механизмы, связывающие материнскую среду с неблагоприятными краткосрочными и долгосрочными последствиями, остаются плохо изученными. В данном обзоре представлены современные знания о влиянии материнского ожирения во время беременности на ребенка в первый год жизни. Понимание этих процессов имеет ключевое значение для разработки терапевтических вмешательств с целью предотвращения будущих сердечно-сосудистых и метаболических патологий у последующих поколений.

Ключевые слова: материнское ожирение; перинатальный период; метаболический синдром; потомство матерей с ожирением; мать-плацента-плод.

INTRODUCTION

Obesity is a serious medical and social problem in modern medicine, it reaches epidemic proportions worldwide. In 2022, according to the World Health Organization report on obesity in Europe, about 55.5% of adult population is overweight and obese. The prevalence of obesity in women of childbearing age is steadily increasing [1]. In 2017–2018, the prevalence of obesity in women of childbearing age in the US was about 40% [2–4], in the UK, 21.6% of women had obesity and 27.4% were overweight. In Scotland in 2021, 25.9% of women were obese by the time of pregnancy [3]. Statistical data in the Russian Federation echo the global trend. Thus, according to the results of the epidemiologic study ESSE-RF (Epidemiology of cardiovascular diseases and their risk factors in the regions of the Russian Federation) in 2013–2014, obesity was found in almost every third citizen of Russia. The second wave of the ESSE-RF program, conducted in 2017, included 17 regions and more than 26,000 participants of both sexes aged from 20 to 64 years. The results of the study showed that obesity was registered in 27.9% of men and 31.8% of women, the prevalence of obesity in women of childbearing age reached 25% [5]. Currently, the problem of obesity in women of childbearing age is urgent, according to recent studies demonstrating the adverse effect of maternal obesity on the health of offspring. Obesity significantly complicates the course of pregnancy and labor, contributing to obstetric complications 2–3 times more often than in women with normal body mass index [6–8]. In addition, women with obesity are more likely than women with normal body mass index (BMI) to have excessive gestational weight gain, which has also been shown to increase obstetric and perinatal risks [9–12].

THE IMPACT OF MATERNAL OBESITY ON FETAL GROWTH AND DEVELOPMENT. PERINATAL RISKS

Numerous studies have proven the role of maternal obesity in the formation of various complications of pregnancy and childbirth, such as pregnancy failure, preeclampsia, gestational diabetes mellitus, gestational arterial hypertension, labor anomalies, increased incidence of operative delivery, bleeding in labor and early postpartum period, maternal and fetal traumas, surgical infections, and delayed fetal intrauterine development [14–18]. In addition to complications of pregnancy and labor, perinatal risks, the mechanisms of long-term effects of maternal obesity on offspring are actively studied. Currently, there are sporadic

studies investigating the impact of maternal obesity in the first year of a child's life, the influence of genetic and epigenetic factors.

During pregnancy, significant anatomical and physiological changes occur in many organs and systems of a woman's body to ensure nutrition and development of the fetus. One of the important mechanisms of physiologic adaptation of the maternal body to pregnancy is the controlled production of cytokines, inflammatory and proinflammatory factors by different cell subtypes at the maternal-fetal interface, since strict regulation of inflammatory factors is required for implantation, placentation, and continuation of pregnancy [19, 20]. Maternal obesity is associated with changes in the profile of the inflammatory response, which directly affects physiologic adaptation. Pregnancy with obesity and excessive body weight results in low-grade chronic inflammation secondary to an impaired immune cell profile, subsequently leading to activation of pro-inflammatory mechanisms. This condition has the name "metaflammation" in the current literature. Nowadays, metaflammation is recognized as a major factor affecting offspring health in early life [21, 22]. Three immunological stages based on the body's inflammatory response during pregnancy have been described [20, 23]. In the first trimester, the initial pro-inflammatory stage is important for implantation and placentation. With the onset of the II trimester, Th2-type anti-inflammatory and immune stage appears, which is necessary for fetal growth. In the III trimester, the pro-inflammatory stage and Th1-type immune activation initiate labor and delivery [20–23]. In addition to the important role of Th1 and Th2 cells during pregnancy, other T-helper cells such as T-helper 17 (Th17), T-helper 22 (Th22), follicular T-helper (Tfh) and regulatory T-cells (Treg) of the mother and fetus contribute to the continuation of a healthy pregnancy. Th17 and Th22 cells are involved in the induction of immunity against extracellular pathogens at the maternal-fetal interface [20–22]. Uncontrolled Th1 and Th17 response is associated with implantation failure and pregnancy failure [21–28]. Treg cells enhance fetal immune tolerance by suppressing excessive Th1 and Th17 activity and autoimmune response [29]. Follicular T-helper cells in the third trimester provide humoral immunity by activating B-cells to initiate an antibody response outside the follicular and germinal center [30]. Type 1 (Th1) and type 2 (Th2) T-helper cells represent the two major subsets of CD4 T-helper cells that regulate the adaptive immune response [31]. Th1 cells produce high levels of interferon- γ (IFN- γ), interleukin-2 (IL-2), tu-

mor necrosis factor (TNF) and are responsible for phagocyte-dependent inflammation as well as for defense against intracellular pathogens [31]. They also play an important role in the development of organ-specific autoimmune diseases and chronic inflammatory diseases [31]. Th2 cells produce IL-4, IL-5, IL-6, IL-9, IL-10 and IL-13, which leads to an excessive immune response by switching B cells, activating eosinophils and inhibiting phagocytic activity [32]. Despite some inconsistent results, a lot of studies, found in the current literature, show that mothers with obesity before pregnancy have increased levels of pro-inflammatory cytokines such as IL-8, IL-6, CRP, TNF- α and IFN- γ and altered levels of adipokines [33–40]. Inflammation occurring against a background of maternal obesity leads to impaired placental development, which affects both maternal and placental inflammatory profiles [41–43]. Nowadays, the influence of cytokines on implantation and remodeling of spiral arteries has been proven. For example, abnormally high levels of TNF- α can lead to impaired remodeling of spiral arteries [44], and IL-6 increases trophoblast cell migration and invasion, while TNF- α decreases it [45–48]. Recent studies have demonstrated that maternal obesity is associated with increased placental mass and decreased placental efficiency, indicating an adaptation to increased nutrient availability to regulate fetal growth [49–54]. A linear correlation between placental mass and birth weight has also been found [49, 54]. Placental transport has a significant influence on the fetal intrauterine environment [55]. In pregnancies with a background of obesity, abnormal placental vasculature is the most common placental pathology [53, 56–59]. Placental vascular growth is regulated by angiogenic factors including VEGF, placental growth factor (PlGF) forming growth factor- β (TGF- β) and leptin, as well as anti-angiogenic factors such as soluble fms-like tyrosine kinase-1 (sFlt-1) and soluble endoglin (sEng) [7–9, 51]. By the end of the second trimester, the villous vessels begin to form loops and wriggle, dramatically increasing the surface area for nutrient and gas exchange [60]. The imbalance between pro- and anti-angiogenic factors is considered an important link in the pathogenesis of preeclampsia and intrauterine developmental delay [65]. Obesity has been proven to be associated with increased placental expression of VEGF, decreased levels of circulating PlGF and sFlt-1 [66, 67]. Thus, maternal obesity impairs the development of placental architecture, potentially jeopardizing fetal growth and survival [60].

In the early stages of physiologically normal pregnancy, insulin sensitivity is increased, which

promotes glucose uptake by adipose tissue, which in turn prepares the body for increased energy requirements later in pregnancy [61]. Obese women have 50–60% higher postprandial insulin concentrations than women with normal body mass in both early and late pregnancy [62]. Women with obesity also have greater glucose tolerance compared to pregnant women with normal BMI, as evidenced by higher fasting glucose levels 1 and 2 hours after the oral glucose tolerance test (OGTT) [62]. Although glycemic values may not meet the criteria for gestational diabetes mellitus (GDM), an abnormal response to the OGTT in obese women is associated with the risk of delivering a fetus large for gestational age [63]. In addition, in obese or overweight pregnant women, increased levels of circulating cytokines in the maternal blood, such as TNF- α and IL-6, have been reported. This association has been proven in the development of insulin resistance in the first and second trimesters of pregnancy [64–66].

In studies carried out on animal models and then repeated in humans, it was found that maternal obesity leads to decreased transport of oleic acid in the placenta in male children, which is associated with decreased expression of the CD36 transporter (fatty acid translocase) and intracellular fatty acid binding protein (FABP 5) [67]. Increased lipid transfer to the fetus contributes to the development of adipose tissue, and thus the risk of developing overweight offspring [68]. Hyperlipidemia and vascular dysfunction may be an important mediator of cardiometabolic diseases observed in offspring born to obese mothers. Considering all the above mechanisms, it can be concluded that maternal obesity has deleterious effects on offspring health. Short-term adverse fetal outcomes in infants of obese mothers include increased risk of fetal overgrowth, stillbirth, and neonatal hypoglycemia [69, 70]. A meta-analysis of published data from 38 cohorts showed that maternal obesity and even small increases in BMI were associated with an increased risk of intrauterine and infant death. For women with BMI >30 kg/m², the absolute risk per 10,000 pregnancies was 102 and 43 intrauterine and infant deaths, respectively [71]. Pregnancies of overweight women are associated with a 2–3-fold increased risk of fetal macrosomia; this is associated with an increase in absolute fetal size and its fat mass [72–74]. Some studies suggest that the programming of obesity in offspring by maternal obesity may be partially sex-specific, basically, male offspring have a greater susceptibility to the risks of developing of obesity compared to female offspring born to obese mothers

[75]. Fetal overgrowth is the main reason for the increased incidence of cesarean section [76, 77]. Severe neonatal hypoglycemia occurs in 10–15% of neonates and can lead to nervous system damage [78]. In maternal obesity, neonatal hypoglycemia is usually transient and occurs because of inadequate, persistent hyperinsulinemia caused by higher than normal concentrations of glucose in the womb [79].

REGULATORY FACTORS AT THE LEVEL OF MOTHER-PLACENTA-FETUS IN MATERNAL OBESITY

Data from recent studies demonstrate that adipose tissue is an important endocrine organ involved in metabolism through several mechanisms, the most important of which is the secretion of bioactive mediators by adipocytes and other cells [80–88]. These bioactive substances, collectively referred to as "adipokines", are important in the pathophysiology of insulin resistance, hyperlipidemia, inflammation, and metabolic syndrome [89–100]. Metabolic adaptation begins early in pregnancy and is accompanied by changes in maternal hormone production, including prolactin, estrogen, progesterone, and cortisol [101, 102]. Placental hormone secretion, which begins immediately after implantation and continues all pregnancy, is important for maternal metabolic adaptation through indirect modeling of endocrine axes and direct changes in maternal metabolic systems [103].

Glucose metabolism. Glucose is a major substrate for placental and fetal energy metabolism, and normal pregnancy causes marked changes in maternal glucose metabolism, including insulin resistance, activation of hepatic glucose production, and increased insulin release by pancreatic β -cells with higher plasma C-peptide levels; these mechanisms contribute to placental and fetal glucose delivery [61]. Early in gestation, fasting glycaemia levels decrease (compared with pre-pregnancy glucose levels), in part due to hemodilution associated with an increase in maternal circulating blood volume. Maternal fasting glycaemia remains consistently low in the second trimester and reaches even lower values in the third trimester: this is due to increased glucose utilization by the fetal-placental complex [62]. Maternal fasting hypoglycemia during pregnancy is compensated for by increased hepatic gluconeogenesis, which contributes to elevated blood glucose levels and helps maintain nutrient supply to the fetus [63–69]. In contrast, postprandial glucose levels progressively increase during pregnancy compared

to pre-pregnancy levels [106, 107]. This is due to impaired peripheral tissue sensitivity to insulin and hence decreased maternal glucose utilization after meals [108]. Obese women have a higher glucose profile than women with normal BMI [70–73]. Maternal glycaemia is a strong determinant of fetal growth, as evidenced by the continuous association of maternal glucose levels with increasing birth weight [65–67].

Insulin. Early in pregnancy, the mother has increased secretin levels and insulin sensitivity, which stimulates lipogenesis and decreases fatty acid oxidation, causing maternal fat accumulation. In mid-pregnancy, insulin resistance develops to direct available nutrients for fetal growth and development. This state of insulin resistance is exacerbated in pregnant women with a background of GDM or obesity [68–71]. Insulin resistance in obesity leads to maternal hyperglycemia and, consequently, fetal hyperglycemia, because glucose freely passes through the placenta [72–78]. It is already known, that insulin plays a neurotrophic role for many brain regions; it inhibits neuronal apoptosis through activation of protein kinase B and protein kinase C, resulting in increased neuronal survival [79–85]. High insulin levels are essential for proper brain maturation [86]. However, chronic hyperinsulinemia, which is proven to be more common in obese mothers, corresponds to high fetal insulin levels, which contributes to fetal insulin resistance [87–94]. Maternal obesity is also associated with characteristic changes in the release of adipokines, which have systemic effects on metabolism and energy homeostasis [95–98].

Adiponectin. Adiponectin is one of the adipokines produced by adipocytes and the most abundant product of adipose tissue and accounts for 0.01% of total plasma proteins. It plays an important role in the relationship between adipose tissue and other metabolic tissues such as liver and skeletal muscle [45–47]. Adiponectin suppresses hepatic gluconeogenesis and contributes to insulin sensitization [45–59]. As opposed to other adipokines, although it is secreted by adipocytes and its plasma concentration is inversely correlated with BMI [56, 99–102]. During pregnancy, adiponectin levels decrease as the insulin resistance develops in pregnant women, which contributes to decreased glucose uptake and increased lipolysis, moving nutrients such as glucose and lipids to the fetus [103–109]. Studies in mice have shown that in maternal obesity, fetal adiponectin enhances fetal fat deposition, thereby increasing fetal body weight, proving the important role of adiponectin in the regulation of maternal meta-

bolism, placental function, and fetal development [110–114].

Leptin. Leptin is released from adipose tissue in proportion to its mass. Leptin levels increase throughout pregnancy, reaching a maximum level in the third trimester. Overweight or obese mothers have higher levels of serum leptin before pregnancy, so throughout pregnancy, leptin concentrations are higher in mother and fetus compared to mothers with normal BMI [85, 87]. Leptin involved in the development of the nervous system as it is an important trophic factor. In studies, leptin has been shown to bind to the receptors of the satiety center in the hypothalamus and form a negative feedback loop, suppressing increased food intake and thus preventing obesity [36]. It is shown in studies that have been carried out on animal models that obesity is associated with hyperleptinemia in both females and their offspring, and maternal obesity leads to the formation of resistance to leptin and, consequently, the inability of leptin to cause anorexic effects [92].

Ghrelin. Ghrelin is a gut hormone with a strong orexigenic signal. After entering the bloodstream, ghrelin circulates in two major forms: acyl ghrelin and des-acyl ghrelin. The maternal concentration of total ghrelin decreases slightly throughout pregnancy, and there is a positive correlation between the ratio of acylated to total circulating ghrelin in mothers in the third trimester of pregnancy and the baby's birth weight [106–110]. Levels of total ghrelin in cord blood are inversely correlated with neonatal birth weight [115–117]. Studies in animal models and humans have shown that maternal ghrelin regulates fetal development in late pregnancy. Administration of ghrelin to mice during the last week of gestation caused a 10–20% increase in body weight in the offspring [118]. In studies on mice, it was shown that ghrelin has an inhibitory role in the development of neural connections of hypothalamus, acting as a "break" in the balance of the neurotrophic action of leptin, and, therefore, proper expression of ghrelin in the neonatal period is important for children and in older age.

Pre-adipocyte factor. Pre-adipocyte factor (PREF-1) is a secreted protein that inhibits adipocyte differentiation both in vitro and in vivo. Pre-adipocyte factor is synthesized as a transmembrane protein whose ectodomain containing repeats of epidermal growth factor, is cleaved by tumor necrosis factor- α -converting enzyme to release a biologically active soluble form [104–108, 118]. The importance of PREF-1 in adipogenesis has been demonstrated in animal models. Mice

experimentally deprived of pre-adipocyte factor had growth retardation, skeletal abnormalities, tendency to obesity, impaired insulin sensitivity and decreased glucose tolerance, which confirms the role of PREF-1 in the regulation of adipocyte differentiation [13]. During embryonic development, PREF-1 is widely expressed in numerous embryonic tissues: multipotent mesenchymal stem cells, pancreatic glandular cells, ovarian and male glandular cells, and is also involved in the differentiation of the central nervous system, hepatocytes, respiratory epithelial cells, mesodermal cells of the renal proximal tubule, and adrenal cortex [119–121]. Increased levels of PREF-1 are detected in serum, urine, and amniotic fluid during the second trimester of pregnancy [53]. After birth, PREF-1 expression ceases in most tissues and is observed in a limited number of cells: preadipocytes, pancreatic islet cells, thymus stromal cells, and adrenal cortex cells [28, 97].

Growth hormone. Growth hormone is well known for its function in stimulating cell growth, reproduction, and regeneration, so it is extremely important for development. Recent research findings suggest that the brain is an important target for growth hormone in the regulation of food intake, energy expenditure and glycaemia, especially in response to various forms of metabolic stress such as glucoprivation, food restriction and exercise [122–125]. During pregnancy, growth hormone action is associated with the regulation of maternal food intake, insulin and leptin sensitivity, suggesting that growth hormone and other gestational hormones are important in preparing the maternal body for the metabolic needs of the offspring [126]. Currently, little is known about the programming effects of maternal and/or fetal growth hormone on hypothalamic development in the offspring. There is some evidence that growth hormone regulates hypothalamic neurocircuits that control energy homeostasis [59].

Pro- and anti-inflammatory cytokines. Numerous studies show that maternal obesity further increases concentrations of pro-inflammatory cytokines such as IL-6, TNF- α , monocyte chemoattractant protein 1 (MCP-1), IL-8 and C-reactive protein in plasma, supporting the concept that the low pro-inflammatory state associated with normal pregnancy is exacerbated by maternal obesity [127]. The biological effects of pro-inflammatory cytokines are counterbalanced by anti-inflammatory cytokines such as IL-1, IL-4, IL-6, IL-10, IL-11 and IL-22 [63]. Obesity is now considered as a key factor in the development of chronic inflammation [96, 128], which is important in the pathogenesis

of pre-eclampsia, gestational diabetes mellitus [129]. Chronic inflammation has unfavorable significance for fetal programming. In a recent animal study, it was shown that the offspring of rats injected with IL-6 throughout pregnancy had more body fat compared to the control group, and male offspring had reduced insulin sensitivity [130].

Lipids. During pregnancy, in maternal organism, a lipid accumulation occurs in the I and II trimesters, and subsequently there is an increase in adipose tissue lipolysis. The catabolic state of female adipose tissue in late pregnancy is associated with hyperlipidemia, mainly corresponding to an increase in plasma triglyceride levels, and a smaller increase in phospholipid and cholesterol levels [70]. Maternal obesity is associated with increased lipid levels, higher triglyceride and very-low-density lipoprotein (VLDL) levels and lower cholesterol, high-density lipoprotein (HDL) levels than women with normal BMI [131]. Several recent studies have shown that maternal postprandial triglycerides and free fatty acids are stronger predictors of neonatal weight gain than maternal glucose levels in obese pregnancies [45]. The amount and nature of fatty acid intake during pregnancy are important for brain development and hypothalamic function in the offspring. Hypothalamic dysfunction was observed in the offspring of mice and rats born to animals that consumed increased amounts of saturated or trans fatty acids [132, 133]. Excessive nutrition usually activates hypothalamic inflammatory signaling through increased endoplasmic reticulum stress in the hypothalamus, which serves as a mechanism for the energy imbalance underlying obesity [76]. In obese pregnant women, offspring have increased level of inflammation in the hypothalamus [58] due to elevated levels of circulating fatty acids [134–138]. It has been suggested that fatty acids play an important role in the hypothalamic dysfunction observed in offspring born to mothers who consume increased amounts of saturated or trans fatty acids, and the mechanisms underlying these changes may be related to endoplasmic reticulum stress and hypothalamic inflammation.

Nowadays, the role of brain-derived neurotrophic factor and peptide YY in regulation at the maternal-placenta-fetal level in maternal obesity and their further effects on child growth and development remain poorly understood [102, 113, 139–141].

THE IMPACT OF MATERNAL OBESITY ON OFFSPRING HEALTH

In addition to adverse intrapartum and perinatal outcomes, maternal obesity is also associated

with the development of chronic diseases in children later in their life. In 1990, David Barker proposed an "adult disease origins" model in which he hypothesized that exposure to a suboptimal environment early in life shapes an individual's future health [80]. Initially, he showed that adults born with low birth weight secondary to intrauterine developmental delay due to inadequate nutrient intake were at higher risk of developing metabolic and cardiovascular diseases. Conversely, conditions associated with intrauterine "overeating" and increased inflammation, such as gestational diabetes mellitus and maternal obesity, negatively impact the long-term health of the offspring. Evidence from recent studies suggests that activation of the proinflammatory state during pregnancy is associated with long-term offspring diseases, including childhood obesity, neuropsychiatric disorders, and allergic diseases [135].

Neuropsychiatric disorders. In recent years, there is increasing evidence that children born to obese mothers are more prone to neurodevelopmental and neuropsychiatric disorders. There is evidence that children born to obese mothers have lower intelligence quotient (IQ), higher rates of autism spectrum disorders (ASD), attention deficit hyperactivity disorder (ADHD), cerebral palsy (CP), and mood disorders [45, 47, 136–138]. In studies on animal models, maternal obesity more often led to neuropsychiatric diseases in offspring, which is associated with significant changes in brain structure in the form of decreased proliferation of neural precursors in the hippocampus, reduced apoptosis in the hippocampus and neuronal differentiation in the dentate gyrus, atrophy of dendrites in the hippocampus and amygdala, and reduced myelination in the cerebral cortex in offspring (predominantly male offspring) [205–209]. It was also shown that offspring born to obese mothers had problems in education, behavioral disorders in the form of hyperactivity, anxiety, decreased sociability, addictive behavior, and food intake disorders [139]. Edlow et al. in their study found that the offspring of mice born to obese animals had increased production of TNF- α in response to polysaccharide exposure in placental CD11b cells compared to control group [140]. The increase in pro-inflammatory cytokines was more significant in male offspring, which may correlate with the prevalence of some neuropsychiatric diseases associated with maternal obesity in males [140–142]. From these findings, it has been hypothesized that it is neuroinflammation and oxidative stress, which arise through increased expression of proinflammatory cytokines,

increased lipid peroxidation and microglia activation in offspring born to obese mothers, that play an important role in adverse neurodevelopmental outcomes [26]. The pro-inflammatory environment affects the metabolism of brain-derived neurotrophic factor (BDNF), which is essential for hippocampal neurogenesis. Alterations in BDNF metabolism and tryptophan hydroxylase (TPH2) expression are associated with anxiety disorder in adulthood [29, 130].

Impact on the immune system. There is evidence that maternal obesity and chronic inflammation during pregnancy increase the risk of developing different chronic diseases in offspring. A lot of such diseases have features of altered immune/inflammatory activation [143–145]. Reviews in recent years have provided evidence of altered fetal immunity in response to maternal obesity [36–38]. In a study conducted by Enninga et al. an increased number of CD4+ lymphocytes and decreased levels of IL-12p40 and chemokines were detected in the cord blood of infants born to obese mothers [146]. In another small cohort study, cord blood from the placenta of obese mothers showed increased numbers of CD3+, CD4+, CD8+, NK and CD8 + CD25 + Foxp3 + Treg lymphocyte subpopulations, while CD34 cells were decreased [40]. Moreover, the effect of maternal obesity on decreased response of fetal monocytes and dendritic cells to toll-like receptor ligands has been proved [147]. The toll-like receptor family plays a key role in the proinflammatory response to bacterial infections; consequently, dysregulation of toll-like receptor signaling is associated with bacterial diseases, including necrotizing enterocolitis [148]. Most of these studies are performed using circulating immune cells and probably do not reflect the specificity of immune cells in different organ systems. Kamimae-Lanning investigated the effect of maternal obesity on hematopoietic stem cells (HSCs) and progenitor cells isolated from the liver of fetal mice [149]. The results showed that female mice that were continuously fed a high-fat diet exhibited not only signs of adverse fetal programming, including growth restriction, but also a decrease in HSCs and progenitor cells in the fetal liver. Despite the decrease in the total number of HSCs and progenitor cells, the proportion of B220+ lymphoid and Gr1+/Ter119+ myeloid cells in the liver was increased, indicating a tendency toward myeloid and B-cell differentiation [43].

Atopic diseases. Several studies and meta-analyses show that children born to obese mothers are at higher risk of developing atopic diseases, including atopic dermatitis and bron-

chial asthma [44–51]. Probably, this is due to altered function of immune checkpoints in offspring born to obese mothers that regulate development of allergy. Elevated levels of maternal TNF- α and pro-inflammatory cytokines, which are significantly elevated in obese mothers, have been associated with frequent lower respiratory tract infections and wheezing in offspring [52, 150]. In animal models, MacDonald et al. showed that the contents of bronchoalveolar lavage in offspring born to obese mice had an increased percentage of neutrophils and an increased concentration of IL-6, which allowed them to propose a theory about the role of the influence of an active proinflammatory state in reactive respiratory diseases in children born to obese mothers [151]. In the same study, increased bronchial hyperreactivity was observed during methacholine provocation test [34]. In 2019, Smoothy et al. found increased concentrations of Th1 (TNF- α) and Th2 (IL-5, IL-33) cytokines in bronchoalveolar lavage of obese mice, without any neutrophilia or eosinophilia, and it was hypothesized that these mice are more prone to hyperreactivity to further exposure to allergens or exposure to viruses [152]. Another study found that offspring born to obese mice exhibited an enhanced sensitization reaction in response to allergen (ovalbumin) administration, which was characterized by overproduction of IL-4, IL-13, TNF- α and TGF- β 1 [153]. The same study demonstrated that mice born to obese mothers exhibited increased eosinophilic/neutrophilic infiltration in the parenchyma of lungs, increased collagen deposition and increased mucus hypersecretion [56]. A study conducted by Castro-Rodriguez et al. in 2020 showed an association of high levels of leptin in cord blood with a higher risk of bronchial asthma in children aged 3 years born to obese mothers [154].

Cardiometabolic diseases. The effect of maternal obesity on the risk of cardiometabolic disease in offspring in childhood and adult life has been demonstrated in human and animal models [58, 155]. Cardiometabolic diseases are a complex phenotype of cardiovascular and metabolic dysfunction characterized by insulin resistance, impaired glucose tolerance, dyslipidaemia, obesity, arterial hypertension, and cardiovascular diseases. Recent studies have demonstrated that children born to obese mothers are at higher risk of future cardiovascular disease (excluding congenital heart disease) [155], and a positive correlation was found between maternal pre-pregnancy BMI and increased blood pressure in the child, regardless of the child's BMI [23, 156]. In addition to cardio-

vascular diseases, children born to obese mothers are susceptible to developing of obesity at early age. Evidence from observational studies of mothers and their children in Europe, North America and Australia showed that high maternal BMI before pregnancy and increased body weight during pregnancy were associated with an increased risk of overweight and obesity in offspring throughout childhood [26]. Recent studies have demonstrated the important role of levels of adipokine and leptin in the formation of obesity in children and adults. Obesity is associated with a state of hyperleptinemia and decreased tissue sensitivity to leptin, which subsequently leads to impaired regulation of energy homeostasis [69]. Leptin is secreted into the blood by adipocytes, regulates appetite, metabolism and energy homeostasis, and increases insulin secretion by pancreatic β -cells [157, 158]. During pregnancy, leptin levels regulate fetal development and growth and are positively correlated with neonatal body weight and fat mass [71–75]. Several studies have shown that low levels of cord blood leptin in children predict increased body weight and body length at 2–3 years of age [159]. Leptin and insulin control metabolism of glucose by acting at peripheral and central units [160]. Insulin is a key regulator of leptin metabolism; hyperinsulinemia leads to an increase in serum leptin concentrations [144–159]. Such infants were more resistant to insulin with a positive correlation with neonatal fat deposition [42, 94]. In the view of fact, that leptin and insulin are factors that influence the development of hypothalamus, nervous system, and involved in appetite regulation. It is shown that maternal obesity programs obesity in their offspring with occurrence of hyperphagia. The hyperphagia has been observed in several different rodent models of maternal obesity in both male and female offspring [161]. Considered, that this hyperphagic phenotype may be caused by altered development and function of hypothalamic circuits that regulate appetite and energy expenditure. The timing of the maximum level of neonatal leptin in rodents is a critical window for the development and maturation of hypothalamic neural connections, because the correct levels and timing of influences are required for proper hypothalamic development. Thus, any influences that result in under- or over-exposure to leptin during these critical periods of development may have negative consequences. In animal models, newborn rats born and obese animals have been shown to have an enhanced and prolonged postnatal leptin surge [162]. Studies both in human and animals show that maternal obesity has sex-specific

effects on glucose metabolism and cardiometabolic profiles in male offspring [163]. One theory is differences in pancreatic β -cell function that are partially associated with increased oxidative stress in the islets of Langerhans and decreased plasma estradiol levels in male offspring. Maternal obesity induces insulin resistance and impairs pancreatic β -cell function, accompanied by inflammation in adipose tissue and hepatic steatosis with marked sex differences [91]. Estrogen in female offspring may play a protective role against oxidative stress induced by the effects of maternal obesity [164].

Diseases of urinary system. Recent studies have shown an association between maternal obesity and congenital abnormalities of the urinary system and reduced volume of fetal kidney in late pregnancy compared with fetal body weight [93–95]. Since kidney volume is proposed as an approximate measure of the number of nephrons, maternal obesity may be associated with a reduction in the number of fetal nephrons, potentially leading to hyperfiltration with further development of chronic kidney disease [165, 166]. In contrast, a study in animal models (rats) showed no effect on the number of fetal nephrons late in pregnancy in obese females [108]. However, there was evidence of increased cellular stress, inflammation and apoptosis in the kidneys of fetuses of obese females [167]. In the postnatal period, studies in rodent have shown that offspring from obese mothers show abnormalities in kidney structure due to oxidative stress and fibrosis [99–102, 168, 169]. A potential mechanism for programming renal dysfunction in offspring is the depression of sirtuin 1 (SIRT1) expression induced by maternal obesity [170, 171]. Sirtuin 1 is a key regulator that promotes lipid utilization and suppresses lipogenesis. It's well known, SIRT1 is reduced in cells with high insulin resistance [109, 110]. Maternal obesity during intrauterine development can lead to increased formation of glomerulosclerosis in response to inflammation with further decline in renal function [60].

Features of breastfeeding. Mother's milk realizes the connection between the health of the mother and the offspring. The triad "mother — breast milk — infant" is a unified system, the basic mechanisms of which have not yet been fully elucidated. The nature of nutrition of a pregnant woman significantly affects the development of the fetus, the state of health of the child in the future. Obese women are less likely to initiate breastfeeding than normal-weight women, and are more at risk of lactation difficulties, which may lead to discontinuation of breastfeeding [61].

Lactation function in women with obesity is affected by physiological (delayed lactogenesis or reduced prolactin production in response to suckling) and psychosocial factors [62–64]. The trophic status of a lactating woman affects the composition of breast milk and, consequently, the rate of growth and development of the infant. One of the main sources of energy is lipids. A systematic review of 11,373 publications found a positive correlation between maternal BMI and the amount of fat in breast milk. For every unit of maternal BMI, 0.56 g/L of fat was added to breast milk, and this association was observed from the 1st to the 6th month after delivery. There was no significant association between maternal BMI and the energy value of milk, lactose content and total protein [168, 169]. There is also evidence that the milk of an obese mother contributes to the formation of components of metabolic syndrome in the child in the future life.

MATERNAL OBESITY AND GUT MICROBIOTA IN CHILDREN

The hypothesis that the gut microbiota is an important factor in the pathogenesis of obesity has led to the investigation of its diversity in a group of overweight and obese individuals. The first evidence suggesting a link between the gut microbiota and obesity was suggested by Ley et al. in a study using 16SrRNA genome sequencing. In their work in animal models, they identified the two most abundant types of bacteria, *Firmicutes* (60–80%) and *Bacteroidetes* (20–40%), which differed proportionally in obese mice compared to mice with normal BMI [170]. Specifically, obese mice showed a 50% decrease in the *Bacteroidetes* population and a proportional increase in *Firmicutes*. Of particular interest were the results which revealed that after dietary treatment, the relative abundance of *Bacteroidetes* increased and *Firmicutes* decreased [171–180]. Turnbaugh et al. confirmed the increased ratio of *Firmicutes* and *Bacteroidetes* in obese mice compared to lean mice in animal models using the latest DNA metagenomic sequencing technique [176, 179]. Moreover, obese mice had a higher proportion of archaea in the microbial communities of the caecum [43]. Armougom et al. in their study evaluated the expression profiles of gut microbiota using real-time PCR and found significantly reduced levels of *Bacteroidetes* in obese individuals compared to those with normal BMI, whereas the concentration of *Firmicutes* was similar in the compared groups [173]. Species-specific variations of *Lactobacillus* in patients with obesity, such as *L. reuteri* and *L. Gasseri*, and lower concen-

trations of *Ruminococcus flavefaciens*, a subgroup of *Ruminococcus flavefaciens* belonging to the bacterial subdivision *Firmicutes*, were also observed in obese individuals [45, 146]. In another study investigating the relationship between gut microbiota, genotype and host's weight, Turnbaugh et al. analyzed the composition of the gut microbiota in monozygotic and dizygotic twins with normal BMI and obesity and their mothers [170]. The results showed that obesity was associated with a low proportion of *Bacteroidetes* and a higher proportion of *Actinobacteria* in obese individuals compared to lean people, but no differences in the phylum *Firmicutes* were found between groups [176, 179]. Numerous studies in recent years have focused on the dynamics of changes in the levels of the bacterial types *Bacteroidetes* and *Firmicutes* in people both with obesity and loss of weight, but there are researches linking obesity in mice to specific bacteria, particularly *Halomonas* and *Sphingomonas*, and decreased numbers of *Bifidobacteria* [180]. A special place in the genesis of obesity is also allocated to archaea — *Methanobrevibacter* is the main representative of archaea in the gut microbiota [149]. Zhang and Armougom et al. found higher numbers of *M. smithii* in obese people compared to a group of people with normal BMI [131, 138, 153, 178]. Currently, there are several putative mechanisms that contribute to the development of obesity. The first is that different strains of the gut microbiota are able to induce low-grade inflammation by stimulating the production of pro-inflammatory cytokines [181–196]. Gram-negative bacteria such as *Bacteroidetes* produce lipopolysaccharide (LPS, endotoxin), which is an important component of the cell wall [197–215]. Cani et al. described that a high-fat diet increases LPS levels, and observational studies have reported diurnal fluctuations in plasma LPS concentration, termed "metabolic endotoxemia" [216]. The pattern of weight gains, visceral and subcutaneous obesity in LPS-injected mice was similar to those observed in mice fed a high-fat diet [217]. In addition, "metabolic endotoxemia" triggered the expression of inflammatory cytokines and serum amyloid A (SAA) proteins. Overgrowth of Gram-negative bacteria such as *Veillonella* in obese individuals can lead to a higher dose of LPS in the intestine, consequently it can disrupt the intestinal barrier through activation of the TLR4/MyD88/IRAK4 signaling pathway in intestinal epithelial cells, resulting in the movement of LPS from the intestine into the bloodstream [183, 184]. When circulating systemically, LPS is able to initiate an immune response in adipose tissue and liver. LPS

first binds to lipopolysaccharide-binding protein, forming a complex with CD14, further inducing the expression of activator protein 1 and nuclear factor kappa B (NF- κ B) by activating toll-like receptor 4 (TLR4), expressed on macrophages and adipose tissue, which promotes the secretion of pro-inflammatory cytokines and chemokines including TNF- α , IL-6, and monocyte chemoattractant protein-1 (MCP-1) [218, 219]. These cytokines can influence adipocytes and stimulate cytokine and chemokine secretion by autocrine and paracrine pathways [185–191, 220–224]. Moreover, MCP-1 overexpression in adipose tissue has been shown to be associated with increased macrophage infiltration in rodents [225].

It's known, the gut microbiota is involved in the central modulation of appetite through the production of gut hormones such as peptide YY (PYY), glucagon-like peptide-1 (GLP-1), and neurotransmitters. *Bifidobacterium* and *Lactobacillus* can produce lactate, which serves as a substrate for neuronal cells, thereby prolonging the post-prandial feeling of satiety [226]. Acetate is able to activate the citric acid cycle in the hypothalamus and further alter the expression profile of neuropeptides regulating satiety [194]. Butyrate affects appetite and eating behavior of the host by activating the vagus nerve and hypothalamus, it is able to cross the blood-brain barrier [193]. Bile acids, short-chain fatty acids and indoles are closely related to the secretion of intestinal hormones by neuroendocrine cells [195–198]. GLP-1 and PYY are the potent anorexigenic hormones that can influence host appetite and eating behavior by binding to their receptors locally distributed in intestinal neurons, vagus nerve afferents, hypothalamus and brainstem [51, 199, 227–230]. The gut microbiota also leads to the production of neurotransmitters, including γ -aminobutyric acid (GABA) and serotonin [37]. GABA, as the predominant inhibitory neurotransmitter of the nervous system, has the ability to stimulate appetite, while serotonin contributes to appetite suppression through regulation of melanocortin neurons [200–204].

In addition, the gut microbiota influences the food center and eating behavior through the regulation of mood. On the one hand, gut microbiota is able to alter mood by affecting the production of bacterial metabolites, gut hormones and neurotransmitters that act as important messengers in gut-brain interactions and further regulate host appetite and eating behavior [47, 205]. On the other hand, the gut microbiota is involved in the regulation of mood and reward pathways,

which presumably influence brain circuits related to eating behavior [206–210, 231].

MATERNAL OBESITY IN PREGNANCY AND CHANGES IN GUT MICROBIOTA

The composition of gut microbiota in pregnant women with obesity differs from that of pregnant women with normal BMI. Physiological shifts in the gut microbiota during pregnancy are necessary to adapt the mother to pregnancy and promote optimal fetal growth and development. During pregnancy on the background of obesity, changes in the gut microbiota may lead to metabolic disturbances of mother, which may indirectly affect the growth and development of the child and the establishment of its own gut microbiota [211–218, 232–242]. Collado et al. in their study observed significant differences in microbial composition in pregnant women depending on their BMI. They found higher numbers of *Bacteroides* and *Staphylococcus aureus* in obese women compared to women with normal BMI [187, 243, 244]. Interestingly, the composition of the microbiota varied with weight gain throughout pregnancy: *Bacteroides* showed a positive correlation both with pre-pregnancy BMI and with weight gain during pregnancy; each kilogram of weight gain was proportionally accompanied by an increase in the number of *Bacteroides* by 0.006 logarithmic units [220–235]. Various studies have shown that the gut microbiota remodels and fluctuates during pregnancy depending on gestational age [221, 236–238]. Zacarias et al. demonstrated that pregnant women with obesity have a high ratio of *Firmicutes* and *Bacteroidetes*, increased numbers of actinobacteria in the second and third trimester of pregnancy, and decreased bacterial diversity in the third trimester [239]. Santacruz et al. studied the fecal microbiota of 50 pregnant women (group 1 — overweight pregnant women, group 2 — pregnant women with normal BMI) to evaluate the relationship between changes in the composition of the gut microbiota during pregnancy and biochemical parameters depending on their BMI. It was found that higher concentration of *Staphylococcus* was significantly correlated with increased serum cholesterol levels, higher number of *Enterobacteriaceae* and *E. coli* was correlated with increased serum ferritin and decreased transferrin levels, while higher number of *Bifidobacterium* was correlated with decreased ferritin and increased transferrin and folic acid levels. The number of *Bacteroides* was associated with higher levels of cholesterol, HDL, and folic acid [240].

THE ROLE OF MATERNAL MICROBIOTA IN PROGRAMMING OF BABY'S OBESITY

It is well known that the first microbial influence on the child is exerted by the maternal microbiota during pregnancy, suggesting that the maternal gut microbiota has a direct influence on the child's gut microbiota and subsequent metabolic and immunologic programming. Both animal and human studies have shown that changes in the diversity and abundance of gut microbial composition in obese mothers were associated with changes in the gut microbiota of the offspring at early and later ages. Soderborg et al. showed in their study that germ-free mice colonized with stool microbes from the stools of two-week-old infants born to obese mothers had increased gut permeability, impaired macrophage activity, and increased inflammation compared to mice colonized with stool microbes from infants born to normal weight mothers [241]. In addition, these mice showed accelerated body weight gain at follow-up [242]. When comparing the gut microbiota in obese and normal weight children, studies have demonstrated an increased ratio of *Firmicutes/Bacteroidetes* in the obese group [243, 244]. Recent studies also show a decrease in the level of bifidobacteria in the intestinal microbiota in obese and overweight children [245]. A study on 77 children born to obese mothers and women with normal BMI showed that the number of *Parabacteroides* spp. and *Oscillibacter* spp. in the gut microbiota was higher in children born to obese mothers. In addition, amounts of *Blautia* spp. and *Eubacterium* spp. were lower [245–251]. Vael et al. in a prospective study demonstrated high intestinal concentrations of *Bacteroides fragilis* and low concentrations of *Staphylococcus* in infants aged from three weeks to one year, which is associated with a higher risk of obesity later in life [247, 252]. Nadal et al. found significantly reduced levels of *Clostridium hystolicum*, *Eubacterium rectale* and *Clostridium coccoides* correlated with weight loss in obese adolescents [253].

Changes in the gut microbiota of offspring born to obese mothers are still controversial and require further investigation.

ADDITIONAL INFORMATION

Author contribution. Thereby, all authors made a substantial contribution to the conception of the study, acquisition, analysis, interpretation of data for the work, drafting and revising the article, final approval of the version to be published and agree to be accountable for all aspects of the study.

Competing interests. The authors declare that they have no competing interests.

Funding source. This study was not supported by any external sources of funding.

ДОПОЛНИТЕЛЬНАЯ ИНФОРМАЦИЯ

Вклад авторов. Все авторы внесли существенный вклад в разработку концепции, проведение исследования и подготовку статьи, прочли и одобрили финальную версию перед публикацией.

Конфликт интересов. Авторы декларируют отсутствие явных и потенциальных конфликтов интересов, связанных с публикацией настоящей статьи.

Источник финансирования. Авторы заявляют об отсутствии внешнего финансирования при проведении исследования.

REFERENCES

1. World Health Organization (WHO). Obesity and overweight. January 2015. <http://www.who.int/mediacentre/factsheets/fs311/en/>. Accessed 2 April 2016.
2. Hruby A., Hu F.B. The epidemiology of obesity: a big picture. *Pharmacoeconomics*. 2015; 33: 673–89.
3. Poston L., Caleyachetty R., Cnattingius S. et al. Pre-conceptional and maternal obesity: epidemiology and health consequences. *Lancet Diabetes Endocrinol*. 2016; 4(12): 1025–36. DOI: 10.1016/S2213-8587(16)30217-0.
4. Hales C.M., Carroll M.D., Fryar C.D., Ogden C.L. Prevalence of Obesity and Severe Obesity Among Adults: United State-2018. *NCHS Data Brief*. 2020; 360: 1–8.
5. Gritsinskaya V.L., Novikova V.P., Khavkin A.I. K voprosu ob epidemiologii ozhireniya u detey i podrostkov (sistematicheskii obzor i meta-analiz nauchnykh publikatsiy za 15-letniy period). [On the issue of the epidemiology of obesity in children and adolescents (a systematic review and meta-analysis of scientific publications over a 15-year period)]. *Voprosy prakticheskoy pediatrii*. 2022; 17(2): 126–35. DOI: 10.20953/1817-7646-2022-2-126-135. (in Russian).
6. Gaillard R., Steegers EAP., Franco O.H. et al. Maternal weight gain in different periods of pregnancy and childhood cardio-metabolic outcomes. The Generation R Study. *International Journal of Obesity*. 2015; 39: 677–85.
7. Hinkle S.N., Sharma A.J., Swan D.W. et al. Excess Gestational Weight Gain Is Associated with Child Adiposity among Mothers with Normal and Overweight Prepregnancy Weight Status. *The Journal of Nutrition*. 2012; 142: 1851–8.

8. Hochner H., Friedlander Y., Calderon-Margalit R. et al. Associations of maternal prepregnancy body mass index and gestational weight gain with adult offspring cardiometabolic risk factors: the Jerusalem Perinatal Family Follow-up Study. *Circulation*. 2012; 125: 1381–9.
9. Oken E., Rifas-Shiman S.L., Field A.E. et al. Maternal Gestational Weight Gain and Offspring Weight in Adolescence. 2008; 112: 8.
10. Kaar J.L., Crume T., Brinton J.T. et al. Maternal Obesity, Gestational Weight Gain, and Offspring Adiposity: The Exploring Perinatal Outcomes among Children Study. *The Journal of Pediatrics*. 2014; 165: 509–15.
11. Josey M.J., McCullough L.E., Hoyo C., Williams-DeVane C. Overall gestational weight gain mediates the relationship between maternal and child obesity. *BMC public health*. 2019; 19: 1062.
12. Arrowsmith S., Wray S., Quenby S. Maternal obesity and labour complications following induction of labour in prolonged pregnancy. *BJOG*. 2011; 118(5): 578–88.
13. Yogev Y., Catalano P.M. Pregnancy and obesity. *Obstet. Gynecol. Clin. North Am.* 2009; 36(2): 285–300.
14. Bogaerts A., Witters I., Van den Bergh B.R. et al. Obesity in pregnancy: altered onset and progression of labour. *Midwifery*. 2013; 29(12): 1303–13.
15. HAPO Study Cooperative Research Group, Metzger B.E., Lowe L.P., Dyer A.R., Trimble E.R., Chaovarindr U. et al. Hyperglycemia and adverse pregnancy outcomes. *N Engl J Med*. 2008; 358(19): 1991–2002.
16. Wu D., Hu D., Chen H. et al. Glucose-regulated phosphorylation of TET2 by AMPK reveals a pathway linking diabetes to cancer. *Nature*. 2018; 559: 637–41.
17. Komshilova K.A., Dzgoeva F.Kh. Beremennost' i ozhireniye [Pregnancy and obesity]. *Ozhireniye i metabolism*. 2009; 4: 9–13.
18. Chukhareva N.A., Runikhina N.K., Dudinskaya Ye.N. Osobennosti techeniya beremennosti u zhen-shchin s ozhireniyem. [Features of the course of pregnancy in women with obesity]. *Akusherstvo i ginekologiya*. 2014; 2: 9–13. (in Russian).
19. Lazo-de-la-Vega-Monroy M-L., Mata-Tapia K-A., Garcia-Santillan J-A. et al. Association of placental nutrient sensing pathways with birth weight. *Reproduction*. 2020; 160: 455–68.
20. Zhu J. T Helper Cell Differentiation, Heterogeneity, and Plasticity. *Cold Spring Harb. Perspect. Biol*. 10(10), 1–17. DOI: 10.1101/cshperspect.a030338.
21. Zhu M.J., Du M., Nathanielsz P.W., Ford S.P. Maternal obesity up-regulates inflammatory signaling pathways and enhances cytokine expression in the mid-gestation sheep placenta. *Placenta*. 2010; 31(5): 387–91. DOI: 10.1016/j.placenta.2010.02.002.
22. Romagnani S. T-cell subsets (Th1 versus Th2). *Ann. Allergy Asthma Immunol*. 2000; 85(1): 9–18. DOI: 10.1016/S1081-1206(10)62426-X.
23. Liang T., Jinglong X., Shusheng D., Aiyu W. Maternal obesity stimulates lipotoxicity and up-regulates inflammatory signaling pathways in the full-term swine placenta. *Anim Sci J*. 2018; 89: 1310–22.
24. Brass E., Hanson E., O'Tierney-Ginn P.F. Placental oleic acid uptake is lower in male offspring of obese women. *Placenta*. 2013; 34: 503–9.
25. Jansson N., Rosario F.J., Gaccioli F. et al. Activation of placental mTOR signaling and amino acid transporters in obese women giving birth to large babies. *J Clin Endocrinol Metab*. 2013; 98: 105–13.
26. Flenady V., Koopmans L., Middleton P. et al. Major risk factors for stillbirth in high-income countries: a systematic review and meta-analysis. *Lancet Lond Engl*. 2011; 377(9774): 1331–40.
27. Aune D., Saugstad O.D., Henriksen T., Tonstad S. Maternal body mass index and the risk of fetal death, stillbirth, and infant death: a systematic review and meta-analysis. *JAMA*. 2014; 311: 1536–46.
28. Ehrenberg H.M., Mercer B.M., Catalano P.M. The influence of obesity and diabetes on the prevalence of macrosomia. *Am J Obstet Gynecol*. 2004; 191: 964–8.
29. Sewell M.F., Huston-Presley L., Super D.M. et al. Increased neonatal fat mass, not lean body mass, is associated with maternal obesity. *Am J Obstet Gynecol*. 2006; 195: 1100–3.
30. Whitelaw A.G. Influence of maternal obesity on subcutaneous fat in the newborn. *Br Med J*. 1976; 1: 985–6.
31. Andres A., Hull H.R., Shankar K. et al. Longitudinal body composition of children born to mothers with normal weight, overweight, and obesity. *Obes Silver Spring Md*. 2015; 23: 1252–8.
32. Jolly M.C., Sebire N.J., Harris J.P. et al. Risk factors for macrosomia and its clinical consequences: a study of 350, 311 pregnancies. *Eur J Obstet Gynecol Reprod Biol*. 2003; 111(1): 9–14.
33. Nesbitt T.S., Gilbert W.M., Herrchen B. Shoulder dystocia and associated risk factors with macrosomic infants born in California. *Am J Obstet Gynecol*. 1998; 179(2): 476–80.
34. Turner D., Monthé-Drèze C., Cherkerzian S. et al. Maternal obesity and cesarean section delivery: additional risk factors for neonatal hypoglycemia? *J Perinatol Off J Calif Perinat Assoc*. 2019; 39(8): 1057–64.
35. Stanley C.A., Rozance P.J., Thornton P.S. et al. Re-evaluating “transitional neonatal hypoglyce-

- mia": mechanism and implications for management. *J Pediatr*. 2015; 166(6): 1520–5.e1.
36. Gimeno R.E., Klamann L.D. Adipose tissue as an active endocrine organ: recent advances. 2005; 5: 122–8.
37. Hutley L., Prins J.B. Fat as an endocrine organ: relationship to the metabolic syndrome. 2005; 330: 280–9.
38. Kahn B.B., Flier J.S. Obesity and insulin resistance. 2000; 106: 473–81.
39. Kahn S.E., Hull R.L., Utzschneider K.M. Mechanisms linking obesity to insulin resistance and type 2 diabetes. 2006; 444: 840–6.
40. Matsuzawa Y., Funahashi T., Nakamura T. Molecular mechanism of metabolic syndrome X: contribution of adipocytokines adipocyte-derived bioactive substances. 1999; 892: 146–54.
41. Montague C.T., O'Rahilly S. The perils of portliness: causes and consequences of visceral adiposity. 2000; 49: 883–8.
42. Ronti T., Lupattelli G., Mannarino E. The endocrine function of adipose tissue: an update. 2006; 64: 355–65.
43. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. 2001; 104: 531–43.
44. Trayhurn P. Endocrine and signalling role of adipose tissue: new perspectives on fat. 2005; 184: 285–93.
45. Catalano P.M., Hoegh M., Minium J. et al. Adiponectin in human pregnancy: implications for regulation of glucose and lipid metabolism. 2006; 49: 1677–85.
46. Farvid M.S., Ng T.W., Chan D.C. et al. Association of adiponectin and resistin with adipose tissue compartments, insulin resistance and dyslipidaemia. 2005; 7: 406–13.
47. Gable D.R., Hurel S.J., Humphries S.E. Adiponectin and its gene variants as risk factors for insulin resistance, the metabolic syndrome and cardiovascular disease. 2006; 188: 231–44.
48. Kirwan J.P., Hauguel-De M.S., Lepercq J. et al. TNF-alpha is a predictor of insulin resistance in human pregnancy. 2002; 51: 2207–13.
49. Lopez-Bermejo A., Fernandez-Real J.M., Garrido E. et al. Maternal soluble tumour necrosis factor receptor type 2 (sTNFR2) and adiponectin are both related to blood pressure during gestation and infant's birthweight. 2004; 61: 544–52.
50. Matsuzawa Y. The metabolic syndrome and adipocytokines. 2006; 580: 2917–21.
51. McLachlan K.A., O'Neal D., Jenkins A., Alford F.P. Do adiponectin, TNFalpha, leptin and CRP relate to insulin resistance in pregnancy? Studies in women with and without gestational diabetes, during and after pregnancy. *Diabetes Metab Res Rev*. 2006; 22: 131–8.
52. Ouchi N., Kihara S., Arita Y. et al. Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF-kappaB signaling through a cAMP-dependent pathway. 2000; 102: 1296–1301.
53. Retnakaran R., Hanley A.J., Raif N. et al. Reduced adiponectin concentration in women with gestational diabetes: a potential factor in progression to type 2 diabetes. 2004; 27: 799–800.
54. Silha J.V., Krsek M., Skrha J.V. et al. Plasma resistin, adiponectin and leptin levels in lean and obese subjects: correlations with insulin resistance. 2003; 149: 331–5.
55. Steppan C.M., Lazar M.A. Resistin and obesity-associated insulin resistance. 2002; 13: 18–23.
56. Tilg H., Moschen A.R. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. 2006; 6: 772–83.
57. Unger R.H. Hyperleptinemia: protecting the heart from lipid overload. 2005; 45: 1031–4.
58. Smirnova N.N., Kupriyenko N.B., Petrenko Yu.V., Novikova V.P. Materinskoye ozhireniye i sistema "mat'-platsenta-plod": dokazannyye mekhanizmy vliyaniya. [Maternal obesity and the "mother-placenta-fetus" system: proven mechanisms of influence]. *Children's Medicine of the North-West*. 2021; 9(3): 31–9. (in Russian).
59. Yokota T., Oritani K., Takahashi I. et al. Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. 2000; 96: 1723–32.
60. Harmon K.A., Gerard L., Jensen D.R. et al. Continuous glucose profiles in obese and normal-weight pregnant women on a controlled diet: metabolic determinants of fetal growth. *Diabetes Care*. 2011; 34(10): 2198–204. DOI: 10.2337/dc11-0723.
61. Smirnova N.N., Khavkin A.I., Novikova V.P. Sostav grudnogo moloka pri ozhireniy mater: vliyaniye na razvitiye rebenka. [The composition of breast milk in obese mothers: the impact on the development of the child]. *Voprosy prakticheskoy pediatrii*. 2022; 17(1): 167–76. DOI: 10.20953/1817-7646-2022-1-167-176. (in Russian).
62. Tsepilova M.O., Polyakova K.D. Vliyaniye aktivnykh metabolitov grudnogo moloka i ikh proizvodnykh na organizm novorozhdonnogo. [Influence of active metabolites of breast milk and their derivatives on the body of a newborn]. *Proba pera: Materialy mezhregional'noy nauchnoy konferentsii molodykh uchenykh "VI Malye Aprel'skiye chteniya pamyati professora M.V. Pikkela"*, Arkhangel'sk, 01 aprelya 2023 goda. Vypusk 6. Arkhangel'sk: Severnyy gosudarstvennyy meditsinskiy universitet; 2023. (in Russian).

63. Smirnova N.N., Khavkin A.I., Kupriyenko N.B., Novikova V.P. Bakterii i virusy grudnogo molo-ka. [Bacteria and viruses in breast milk]. Voprosy detskoy diyetologii. 2022; 20(2): 74–82. DOI: 10.20953/1727-5784-2022-2-74-82. EDN BBIKOO. (in Russian).
64. Metzger B.E., Lowe L.P., Dyer A.R. et al. Hyperglycemia and adverse pregnancy outcomes. N Engl J Med. 2008; 358(19): 1991–2002. DOI: 10.1056/NEJMoa0707943.
65. Xiang A.H., Peters R.K., Trigo E. et al. Multiple metabolic defects during late pregnancy in women at high risk for type 2 diabetes. Diabetes. 1999; 48(4): 848–54. DOI: 10.2337/diabetes.48.4.848.
66. Friedman J.E., Ishizuka T., Shao J. et al. Impaired glucose transport and insulin receptor tyrosine phosphorylation in skeletal muscle from obese women with gestational diabetes. Diabetes. 1999; 48(9): 1807–14. DOI: 10.2337/diabetes.48.9.1807.
67. Catalano P.M., Ehrenberg H.M. The short- and long-term implications of maternal obesity on the mother and her offspring. BJOG. 2006; 113(10): 1126–33. DOI: 10.1111/j.1471-0528.2006.00989.x.
68. Catalano P.M., Presley L., Minium J., Hauguel-de Mouzon S. Fetuses of obese mothers develop insulin resistance in utero. Diabetes Care. 2009; 32(6): 1076–80. DOI: 10.2337/dc08-2077.
69. Toran-Allerand C.D., Ellis L., Pfenninger K.H. Estrogen and insulin synergism in neurite growth enhancement in vitro: mediation of steroid effects by interactions with growth factors? Brain Res. 1988; 469(1-2): 87–100. DOI: 10.1016/0165-3806(88)90172-1.
70. Recio-Pinto E., Ishii D.N. Effects of insulin, insulin-like growth factor-II and nerve growth factor on neurite outgrowth in cultured human neuroblastoma cells. Brain Res. 1984; 302(2): 323–34. DOI: 10.1016/0006-8993(84)90246-4.
71. Lázár B.A., Jancsó G., Pálvölgyi L. et al. Insulin confers differing effects on neurite outgrowth in separate populations of cultured dorsal root ganglion neurons: The role of the insulin receptor. Front Neurosci. 2018; 12: 732. DOI: 10.3389/fnins.2018.00732.
72. Song J., Wu L., Chen Z. et al. Axons guided by insulin receptor in drosophila visual system. Science. 2003; 300(5618): 502–5. DOI: 10.1126/science.1081203.
73. Fex Svenningsen A., Kanje M. Insulin and the insulin-like growth factors I and II are mitogenic to cultured rat sciatic nerve segments and stimulate [3H]thymidine incorporation through their respective receptors. Glia. 1996; 18(1): 68–72. DOI: 10.1002/(SICI)1098-1136(199609)18:1.
74. Dudek H., Datta S.R., Franke T.F. et al. Regulation of neuronal survival by the serine-threonine protein kinase akt. Science. 1997; 275(5300): 661–5. DOI: 10.1126/science.275.5300.661.
75. Apostolatos A., Song S., Acosta S. et al. Insulin promotes neuronal survival via the alternatively spliced protein kinase CδII isoform. J Biol Chem. 2012; 287(12): 9299–310. DOI: 10.1074/jbc.M111.313080.
76. Haddad-Tóvolli R., Altirriba J., Obri A. et al. Pro-opiomelanocortin (POMC) neuron translational signatures underlying obesogenic gestational malprogramming in mice. Mol Metab. 2020; 36: 100963. DOI: 10.1016/j.molmet.2020.02.006.
77. Melo A.M., Benatti R.O., Ignacio-Souza L.M. et al. Hypothalamic endoplasmic reticulum stress and insulin resistance in offspring of mice dams fed high-fat diet during pregnancy and lactation. Metabolism. 2014; 63(5): 682–92. DOI: 10.1016/j.metabol.2014.02.002.
78. Wang Z.V., Scherer P.E. Adiponectin, the past two decades. J Mol Cell Biol. 2016; 8: 93–100.
79. Chandran M., Phillips S.A., Ciaraldi T., Henry R.R. Adiponectin: more than just another fat cell hormone? Diabetes Care. 2003; 26: 2442–50.
80. Arita Y., Kihara S., Ouchi N. et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. 1999; 257: 79–83.
81. Hu E., Liang P., Spiegelman B.M. AdipoQ is a novel adipose-specific gene dysregulated in obesity. 1996; 271: 10697–703.
82. Hinkle S.N., Rawal S., Liu D. et al. Maternal adipokines longitudinally measured across pregnancy and their associations with neonatal size, length, and adiposity. Int J Obes (Lond). 2019; 43: 1422–34.
83. Aye I.L., Powell T.L., Jansson T. Review: Adiponectin — the missing link between maternal adiposity, placental transport and fetal growth? Placenta. 2013; 34: S40–5.
84. Qiao L., Yoo H.S., Madon A. et al. Adiponectin enhances mouse fetal fat deposition. Diabetes. 2012; 61: 3199–207.
85. Forhead A.J., Fowden A.L. The hungry fetus? Role of leptin as a nutritional signal before birth. J Physiol. 2009; 587: 1145–52.
86. Pardo I.M., Geloneze B., Tambascia M.A., Barros-Filho A.A. Hyperadiponectinemia in newborns: relationship with leptin levels and birth weight. Obes Res. 2004; 12: 521–4.
87. Frederiksen L., Nielsen T.L., Wraae K. et al. Subcutaneous rather than visceral adipose tissue is associated with adiponectin levels and insulin resistance in young men. J Clin Endocrinol Metab. 2009; 94: 4010–5.

88. Misra V.K., Straughen J.K., Trudeau S. Maternal serum leptin during pregnancy and infant birth weight: the influence of maternal overweight and obesity. *Obes (Silver Spring)*. 2013; 21(5): 1064–9. DOI: 10.1002/oby.20128.
89. Patro-Małyśza J., Trojnar M., Skórzyńska-Dziduszko K.E. et al. Leptin and ghrelin in excessive gestational weight gain-association between mothers and offspring. *Int J Mol Sci*. 2019; 20(10): 2398. DOI: 10.3390/ijms20102398.
90. Trayhurn P., Beattie J.H. Physiological role of adipose tissue: white adipose tissue as an endocrine and secretory organ. *Proc Nutr Soc*. 2001; 60: 329–39.
91. Morrison C.D. Leptin resistance and the response to positive energy balance. *Physiol Behav*. 2008; 94: 660–3.
92. Tham E., Liu J., Innis S. et al. Acylated ghrelin concentrations are markedly decreased during pregnancy in mothers with and without gestational diabetes: relationship with cholinesterase. *Am J Physiol Endocrinol Metab*. 2009; 296(5): E1093–100. DOI: 10.1152/ajpendo.90866.2008.
93. Karakulak M., Saygili U., Temur M. et al. Comparison of umbilical cord ghrelin concentrations in full-term pregnant women with or without gestational diabetes. *Endocr Res*. 2017; 42(2): 79–85. DOI: 10.1080/07435800.2016.1194855.
94. Nakahara K., Nakagawa M., Baba Y. et al. Maternal ghrelin plays an important role in rat fetal development during pregnancy. *Endocrinology*. 2006; 147(3): 1333–42. DOI: 10.1210/en.2005-0708.
95. Wang Y., Sul H.S. Ectodomain shedding of preadipocyte factor 1 (Pref-1) by tumor necrosis factor alpha converting enzyme (TACE) and inhibition of adipocyte differentiation. *Mol Cell Biol*. 2006; 26: 5421–35.
96. Shulman G.I. Cellular mechanisms of insulin resistance. *J Clin Invest*. 2000; 106: 171–6.
97. Garg A. Acquired and inherited lipodystrophies. *N Engl J Med*. 2004; 350: 1220–34.
98. Shackleton S., Lloyd D.J., Jackson S.N. et al. LMNA, encoding lamin A/C, is mutated in partial lipodystrophy. *Nat Genet*. 2000; 24: 153–6.
99. Agarwal A.K., Arioglu E., De Almeida S. et al. AGPAT2 is mutated in congenital generalized lipodystrophy linked to chromosome 9q34. *Nat Genet*. 2002; 31: 21–3.
100. Magre J., Delepine M., Khallouf E. et al. Identification of the gene altered in Berardinelli-Seip congenital lipodystrophy on chromosome 11q13. *Nat Genet*. 2001; 28: 365–70.
101. Barroso I., Gurnell M., Crowley V.E. et al. Dominant negative mutations in human PPARgamma associated with severe insulin resistance, diabetes mellitus and hypertension. *Nature*. 1999; 402: 880–3.
102. Gregoire F.M., Smas C.M., Sul H.S. Understanding adipocyte differentiation. *Physiol Rev*. 1998; 78: 783–809.
103. Rosen E.D., Spiegelman B.M. Molecular regulation of adipogenesis. *Annu Rev Cell Dev Biol*. 2000; 16: 145–71.
104. Smas C.M., Sul H.S. Pref-1, a protein containing EGF-like repeats, inhibits adipocyte differentiation. *Cell*. 1993; 73: 725–34.
105. Bonen A., Parolin M.L., Steinberg G.R. et al. Triacylglycerol accumulation in human obesity and type 2 diabetes is associated with increased rates of skeletal muscle fatty acid transport and increased sarcolemmal FAT/CD36. *Faseb J*. 2004; 18: 1144–6.
106. Smas C.M., Chen L., Sul H.S. Cleavage of membrane-associated pref-1 generates a soluble inhibitor of adipocyte differentiation. *Mol Cell Biol*. 1997; 17: 977–88.
107. Kershaw E.E., Flier J.S. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab*. 2004; 89: 2548–56.
108. Villena J.A., Kim K.H., Sul H.S. Pref-1 and ADSF/resistin: two secreted factors inhibiting adipose tissue development. *Horm Metab Res*. 2002; 34: 664–70.
109. Furigo I.C., Teixeira P.D.S., de Souza G.O. et al. Growth hormone regulates neuroendocrine responses to weight loss via AgRP neurons. *Nat Commun*. 2019; 10(1): 662. DOI: 10.1038/s41467-019-08607-1.
110. Furigo I.C., de Souza G.O., Teixeira P.D.S. et al. Growth hormone enhances the recovery of hypoglycemia. *FASEB J*. 2019; 33(11): 11909–24. DOI: 10.1096/fj.201901315R.
111. Donato J., Wasinski F., Furigo I.C. et al. Central regulation of metabolism by growth hormone. *Cells*. 2021; 10(1): 129. DOI: 10.3390/cells10010129.
112. Teixeira P.D.S., Couto G.C., Furigo I.C. et al. Central growth hormone action regulates metabolism during pregnancy. *Am J Physiol Endocrinol Metab*. 2019; 317(5): E925–E40. DOI: 10.1152/ajpendo.00229.2019.
113. Wasinski F., Furigo I.C., Teixeira P.D.S. et al. Growth hormone receptor deletion reduces the density of axonal projections from hypothalamic arcuate nucleus neurons. *Neuroscience*. 2020; 434: 136–47. DOI: 10.1016/j.neuroscience.2020.03.037.
114. Challier J.C., Basu S., Bintein T. et al. Obesity in pregnancy stimulates macrophage accumulation and inflammation in the placenta. *Placenta*. 2008; 29(3): 274–81.
115. Ramsay J.E., Ferrell W.R., Crawford L. et al. Maternal obesity is associated with dysregulation of

- metabolic, vascular, and inflammatory pathways. *J Clin Endocrinol Metab.* 2002; 87(9): 4231–7.
116. Petrenko Yu.V., Gerasimova K.S., Novikova V.P. Biologicheskaya i patofiziologicheskaya znachimost' adiponektina. [Biological and pathophysiological significance of adiponectin]. *Pediatr.* 2019; 10(2): 83–7. DOI: 10.17816/PED10283-87. EDN RVWXCN. (in Russian).
117. Novikova V.P., Ivanov D.O., Petrenko Yu.V. et al. Increased marker of endothelial cell dysfunction sVCAM-1 in umbilical cord blood in neonates born to obese women. *Archives of Disease in Childhood.* 2019; 104(S3): 117. DOI: 10.1136/archdischild-2019-epa.273.
118. Azizian M., Mahdipour E., Mirhafez S.R. et al. Cytokine profiles in overweight and obese subjects and normal weight individuals matched for age and gender. *Ann Clin Biochem.* 2016; 53(6): 663–8.
119. Brunner S., Schmid D., Hüttinger K. et al. Maternal insulin resistance, triglycerides and cord blood insulin in relation to post-natal weight trajectories and body composition in the offspring up to 2 years. *Diabet Med J Br Diabet Assoc.* 2013; 30(12): 1500–7.
120. Regnault N., Botton J., Heude B. et al. Higher Cord C-Peptide Concentrations Are Associated With Slower Growth Rate in the 1st Year of Life in Girls but Not in Boys. *Diabetes.* 2011; 60(8): 2152–9.
121. Stang J., Huffman L.G. Position of the Academy of Nutrition and Dietetics: Obesity, Reproduction, and Pregnancy Outcomes. *J Acad Nutr Diet.* 2016; 116(4): 677–91.
122. Dubé E., Gravel A., Martin C. et al. Modulation of fatty acid transport and metabolism by maternal obesity in the human full-term placenta. *Biol Reprod.* 2012; 87(1): 14, 1–11.
123. Hellmuth C., Lindsay K.L., Uhl O. et al. Association of maternal prepregnancy BMI with metabolomic profile across gestation. *Int J Obes* 2005. 2017; 41(1): 159–69.
124. Briley A.L., Barr S., Badger S. et al. A complex intervention to improve pregnancy outcome in obese women; the UPBEAT randomised controlled trial. *BMC Pregnancy Childbirth.* 2014; 14: 74.
125. Herrera E., Ortega-Senovilla H. Implications of lipids in neonatal body weight and fat mass in gestational diabetic mothers and non-diabetic controls. *Curr Diabetes Rep.* 2018; 18(2): 7. DOI: 10.1007/s11892-018-0978-4.
126. Merzouk H., Meghelli-Bouchenak M., Loukidi B. et al. Impaired serum lipids and lipoproteins in fetal macrosomia related to maternal obesity. *Biol Neonate.* 2000; 77(1): 17–24. DOI: 10.1159/000014190.
127. Furse S., Koulman A., Ozanne S.E. et al. Altered lipid metabolism in obese women with gestational diabetes and associations with offspring adiposity. *J Clin Endocrinol Metab.* 2022; 107(7): e2825–e32. DOI: 10.1210/clinem/dgac206.
128. Barbour L.A., Farabi S.S., Friedman J.E. et al. Postprandial triglycerides predict newborn fat more strongly than glucose in women with obesity in early pregnancy. *Obes (Silver Spring).* 2018; 26(8): 1347–56. DOI: 10.1002/oby.22246.
129. Pimentel G.D., Lira F.S., Rosa J.C. et al. Intake of trans fatty acids during gestation and lactation leads to hypothalamic inflammation via TLR4/NFκBp65 signaling in adult offspring. *J Nutr Biochem.* 2012; 23(3): 265–71. DOI: 10.1016/j.jnutbio.2010.12.003.
130. Rother E., Kuschewski R., Alcazar M.A. et al. Hypothalamic JNK1 and IKKβ activation and impaired early postnatal glucose metabolism after maternal perinatal high-fat feeding. *Endocrinology.* 2012; 153(2): 770–81. DOI: 10.1210/en.2011-1589.
131. Zhang X., Zhang G., Zhang H. et al. Hypothalamic IKKβ/NF-κB and ER stress link overnutrition to energy imbalance and obesity. *Cell.* 2008; 135(1): 61–73. DOI: 10.1016/j.cell.2008.07.043.
132. Sadagurski M., Debarba L.K., Werneck-de-Castro J.P. et al. Sexual dimorphism in hypothalamic inflammation in the offspring of dams exposed to a diet rich in high fat and branched-chain amino acids. *Am J Physiol Endocrinol Metab.* 2019; 317(3): E526–E34. DOI: 10.1152/ajpendo.00183.2019.
133. Melo A.M., Benatti R.O., Ignacio-Souza L.M. et al. Hypothalamic endoplasmic reticulum stress and insulin resistance in offspring of mice dams fed high-fat diet during pregnancy and lactation. *Metabolism.* 2014; 63(5): 682–92. DOI: 10.1016/j.metabol.2014.02.002.
134. Park S., Jang A., Bouret S.G. Maternal obesity-induced endoplasmic reticulum stress causes metabolic alterations and abnormal hypothalamic development in the offspring. *PloS Biol.* 2020; 18(3): e3000296. DOI: 10.1371/journal.pbio.3000296.
135. Barnes S.K., Ozanne S.E. Pathways linking the early environment to long-term health and lifespan. *Prog Biophys Mol Biol.* 2011; 106(1): 323–36. DOI: 10.1016/j.pbiomolbio.2010.12.005.
136. Drake A.J., Reynolds R.M. Impact of maternal obesity on offspring obesity and cardiometabolic disease risk. *Reproduction.* 2010; 140(3): 387–98. DOI: 10.1530/REP-10-0077.
137. Gaillard R. Maternal obesity during pregnancy and cardiovascular development and disease in the offspring. *Eur J Epidemiol.* 2015; 30(11): 1141–52. DOI: 10.1007/s10654-015-0085-7.
138. Nicholas L.M., Morrison J.L., Rattanatravay L. et al. The early origins of obesity and insulin resistance:

- timing, programming and mechanisms. *Int J Obes (Lond)*. 2016; 40(2): 229–38.
139. Poston L. Developmental programming and diabetes — the human experience and insight from animal models. *Best Pract Res Clin Endocrinol Metab*. 2010; 24(4): 541–52. DOI: 10.1016/j.beem.2010.05.007.
 140. Edlow A.G., Glass R.M., Smith C.J. et al. Placental Macrophages: A Window Into Fetal Microglial Function in Maternal Obesity. *Int. J. Dev. Neurosci*. 2019; 77, 60–8. DOI: 10.1016/j.ijdevneu.2018.11.004.
 141. Khavkin A.I., Ayrumov V.A., Shvedkina N.O., Novikova V.P. Biologicheskaya rol' i klinicheskoye znachenie neuropeptidov v pediatrii: peptid YY i grelin. [Biological role and clinical significance of neuropeptides in pediatrics: peptide YY and ghrelin]. *Voprosy prakticheskoy pediatrii*. 2020; 15(5): 87–92. DOI: 10.20953/1817-7646-2020-5-87-92. (in Russian).
 142. Symonds M.E., Sebert S.P., Hyatt M.A., Budge H. Nutritional programming of the metabolic syndrome. *Nat Rev Endocrinol*. 2009; 5(11): 604–10. DOI: 10.1038/nrendo.2009.195.
 143. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. *Cell*. 2001; 104(4): 531–43. DOI: 10.1016/S0092-8674(01)00240-9.
 144. Hassink S.G., Sheslow D.V., de Lancey E. et al. Serum leptin in children with obesity: relationship to gender and development. *Pediatrics*. 1996; 98(2 Pt 1): 201–3.
 145. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. *Cell*. 2001; 104(4): 531–43. DOI: 10.1016/S0092-8674(01)00240-9.
 146. Morton G.J., Schwartz M.W. Leptin and the central nervous system control of glucose metabolism. *Physiol Rev*. 2011; 91(2): 389–411. DOI: 10.1152/physrev.00007.2010.
 147. Hoggard N., Hunter L., Duncan J.S. et al. Leptin and leptin receptor mRNA and protein expression in the murine fetus and placenta. *Proc Natl Acad Sci U S A*. 1997; 94(20): 11073–8. DOI: 10.1073/pnas.94.20.11073.
 148. Shekhawat P.S., Garland J.S., Shivpuri C. et al. Neonatal cord blood leptin: its relationship to birth weight, body mass index, maternal diabetes, and steroids. *Pediatr Res*. 1998; 43(3): 338–43. DOI: 10.1203/00006450-199803000-00005.
 149. Kamimae-Lanning A.N., Krasnow S.M., Goloviznina N.A. et al. Maternal high-fat diet and obesity compromise fetal hematopoiesis. *Mol. Metab*. 2015; 4(1): 25–38. DOI: 10.1016/j.molmet.2014.11.001.
 150. Boeke C.E., Mantzoros C.S., Hughes M.D. et al. Differential associations of leptin with adiposity across early childhood. *Obesity (Silver Spring)*. 2013; 21(7): 1430–7. DOI: 10.1002/oby.20314.
 151. Mantzoros C.S., Rifas-Shiman S.L., Williams C.J. et al. Cord blood leptin and adiponectin as predictors of adiposity in children at 3 years of age: a prospective cohort study. *Pediatrics*. 2009; 123(2): 682–9. DOI: 10.1542/peds.2008-0343.
 152. Ong K.K., Ahmed M.L., Sherriff A. et al. Cord blood leptin is associated with size at birth and predicts infancy weight gain in humans. *ALSPAC Study Team. Avon Longitudinal Study of Pregnancy and Childhood. J Clin Endocrinol Metab*. 1999; 84(3): 1145–8. DOI: 10.1210/jcem.84.3.5657.
 153. Zuo H.J., Xie Z.M., Zhang W.W. et al. Gut bacteria alteration in obese people and its relationship with gene polymorphism. *World J. Gastroenterol*. 2011; 17: 1076–81. DOI: 10.3748/wjg.v17.i8.1076.
 154. Castro-Rodriguez J.A., Forno E., Casanello P. et al. Leptin in Cord Blood Associates with Asthma Risk at Age 3 in the Offspring of Women with Gestational Obesity. *Ann. Am. Thorac. Soc*. 2020; 17 (12): 1583–9. DOI: 10.1513/AnnalsATS.202001-080OC.
 155. Chang G-Q., Gaysinskaya V., Karatayev O. et al. Maternal High-Fat Diet and Fetal Programming: Increased Proliferation of Hypothalamic Peptide-Producing Neurons That Increase Risk for Overeating and Obesity. *J Neurosci*. 2008; 28: 12107–19.
 156. Nguyen L.T., Saad S., Tan Y. et al. Maternal high-fat diet induces metabolic stress response disorders in offspring hypothalamus. *J Mol Endocrinol*. 2017; 59: 81–92.
 157. Kirk S.L., Samuelsson A-M., Argenton M. et al. Maternal Obesity Induced by Diet in Rats Permanently Influences Central Processes Regulating Food Intake in Offspring. *PLOS ONE*. 2009; 4: e5870.
 158. Glavas M.M., Kirigiti M.A., Xiao X.Q. et al. Early overnutrition results in early-onset arcuate leptin resistance and increased sensitivity to high-fat diet. *Endocrinology*. 2010; 151: 1598–1610.
 159. Long N.M., Ford S.P., Nathanielsz P.W. Maternal obesity eliminates the neonatal lamb plasma leptin peak. *J Physiol*. 2011; 589: 1455–62.
 160. Macumber I., Schwartz S., Leca N. Maternal obesity is associated with congenital anomalies of the kidney and urinary tract in offspring. *Pediatr Nephrol*. 2017; 32: 635–42.
 161. Hsu C.W., Yamamoto K.T., Henry R.K. et al. Prenatal risk factors for childhood CKD. *J Am Soc Nephrol JASN*. 2014; 25: 2105–11.
 162. Lee Y.Q., Lumbers E.R., Oldmeadow C. et al. The relationship between maternal adiposity during pregnancy and fetal kidney development and kidney function in infants: the Gomeri gaaynggal study. *Physiol Rep*. 2019; 7: e14227.

163. Luyckx V.A., Brenner B.M. The clinical importance of nephron mass. *J Am Soc Nephrol JASN*. 2010; 21: 898–910.
164. Brenner B.M., Chertow G.M. Congenital oligonephropathy and the etiology of adult hypertension and progressive renal injury. *Am J Kidney Dis Off J Natl Kidney Found*. 1994; 23: 171–5.
165. Zhou P., Guan H., Guo Y. et al. Maternal High-Fat Diet Programs Renal Peroxisomes and Activates NLRP3 Inflammasome-Mediated Pyroptosis in the Rat Fetus. *J Inflamm Res*. 2021; 14: 5095–5110.
166. Shamseldeen A.M., Ali Eshra M., Ahmed Rashed L. et al. Omega-3 attenuates high fat diet-induced kidney injury of female rats and renal programming of their offsprings. *Arch Physiol Biochem*. 2019; 125: 367–77.
167. Glastras S.J., Chen H., McGrath R.T. et al. Effect of GLP-1 Receptor Activation on Offspring Kidney Health in a Rat Model of Maternal Obesity. *Sci Rep*. 2016; 6: 23525.
168. Glastras S.J., Tsang M. Teh R. et al. Maternal Obesity Promotes Diabetic Nephropathy in Rodent Offspring. *Sci Rep*. 2016; 6: 27769.
169. Yamada-Obara N., Yamagishi S., Taguchi K. et al. Maternal exposure to high-fat and high-fructose diet evokes hypoadiponectinemia and kidney injury in rat offspring. *Clin Exp Nephrol*. 2016; 20: 853–61.
170. Ley R.E., Bäckhed F., Turnbaugh P. et al. Obesity alters gut microbial ecology. *Proc. Natl. Acad. Sci. USA*. 2005; 102: 11070–5. DOI: 10.1073/pnas.0504978102.
171. Nguyen L.T., Mak C.H., Chen H. et al. SIRT1 Attenuates Kidney Disorders in Male Offspring Due to Maternal High-Fat Diet. *Nutrients*. 2019; 11: 146.
172. Jackson C.M., Alexander B.T., Roach L. et al. Exposure to maternal overnutrition and a high-fat diet during early postnatal development increases susceptibility to renal and metabolic injury later in life. *Am J Physiol-Ren Physiol*. 2012; 302: F774–F783.
173. Flynn E.R., Alexander B.T., Lee J. et al. High-fat/fructose feeding during prenatal and postnatal development in female rats increases susceptibility to renal and metabolic injury later in life. *Am J Physiol-Regul Integr Comp Physiol*. 2013; 304: R278–R285.
174. Preveden T., Scarpellini E., Milić N. et al. Gut microbiota changes and chronic hepatitis C virus infection. *Expert Rev. Gastroenterol. Hepatol*. 2017; 11: 813–9. DOI: 10.1080/17474124.2017.1343663.
175. Moore W.E., Holdeman L.V. Human fecal flora: The normal flora of 20 Japanese-Hawaiians. *Appl. Microbiol*. 1974; 27: 961–79.
176. Gill S.R., Pop M., Deboy R.T. et al. Metagenomic analysis of the human distal gut microbiome. *Science*. 2006; 312: 1355–9. DOI: 10.1126/science.1124234.
177. Armougom F., Henry M., Vialettes B. et al. Monitoring bacterial community of human gut microbiota reveals an increase in *Lactobacillus* in obese patients and *Methanogens* in anorexic patients. *PLoS ONE*. 2009; 4: e7125. DOI: 10.1371/journal.pone.0007125.
178. Nash A.K., Auchtung T.A., Wong M.C. et al. The gut mycobiome of the Human Microbiome Project healthy cohort. *Microbiome*. 2017; 5: 153. DOI: 10.1186/s40168-017-0373-4.
179. Turnbaugh P.J., Gordon J.I. The core gut microbiome, energy balance and obesity. *J. Physiol*. 2009; 587: 4153–8. DOI: 10.1113/jphysiol.2009.174136.
180. De Faria Ghetti F., Oliveira D.G., de Oliveira J.M. et al. Influence of gut microbiota on the development and progression of nonalcoholic steatohepatitis. *Eur. J. Nutr*. 2018; 57: 861–76. DOI: 10.1007/s00394-017-1524-x.
181. Ottman N., Smidt H., de Vos W.M., Belzer C. The function of our microbiota: Who is out there and what do they do? *Front. Cell. Infect. Microbiol*. 2012; 2: 104. DOI: 10.3389/fcimb.2012.00104.
182. Sittipo P., Lobionda S., Lee Y.K., Maynard C.L. Intestinal microbiota and the immune system in metabolic diseases. *J. Microbiol*. 2018; 56: 154–62. DOI: 10.1007/s12275-018-7548-y.
183. Koleva P.T., Kim J.S., Scott J.A. and Kozyrskyj A.L. Microbial programming of health and disease starts during fetal life. *Birth Defects Res. C Embryo Today*. 2015; 105: 265–77. DOI: 10.1002/bdrc.21117.
184. Escherich T. The intestinal bacteria of the neonate and breast-fed infant. 1885. *Rev. Infect. Dis*. 1989; 11: 352–6. DOI: 10.1093/clinids/11.2.352.
185. Küstner O. Beitrag zur lehre von der puerperalen infection der neugeborenen. *Archiv. für Gynäkologie*. 1877; 11: 256–63. DOI: 10.1007/BF01845161.
186. Perez-Munoz M.E., Arrieta M.C., Ramer-Tait A.E. and Walter J. A critical assessment of the “sterile womb” and “in utero colonization” hypotheses: Implications for research on the pioneer infant microbiome. *Microbiome*. 2017; 5: 48. DOI: 10.1186/s40168-017-0268-4.
187. Collado M.C., Rautava S., Aakko J. et al. Human gut colonisation may be initiated in utero by distinct microbial communities in the placenta and amniotic fluid. *Sci. Rep*. 2016; 6: 23129. DOI: 10.1038/srep23129.
188. Wassenaar T.M. and Panigrahi P. Is a foetus developing in a sterile environment? *Lett. Appl. Microbiol*. 2014; 59: 572–9. DOI: 10.1111/lam.12334.
189. Jimenez E., Marin M.L., Martin R. et al. Is meconium from healthy newborns actually sterile? *Res. Microbiol*. 2008; 159: 187–93. DOI: 10.1016/j.resmic.2007.12.007.

190. Hu J., Nomura Y., Bashir A. et al. Diversified microbiota of meconium is affected by maternal diabetes status. *PLoS One*. 2013; 8: e78257. DOI: 10.1371/journal.pone.0078257.
191. Zheng J., Xiao X.H., Zhang Q. et al. Correlation of placental microbiota with fetal macrosomia and clinical characteristics in mothers and newborns. *Oncotarget*. 2017; 8: 82314–25.
192. Aagaard K., Ma J., Antony K.M. et al. The placenta harbors a unique microbiome. *Sci. Transl. Med*. 2014; 6: 237ra265.
193. Gomez-Arango L.F., Barrett H.L., McIntyre H.D. et al. Contributions of the maternal oral and gut microbiome to placental microbial colonization in overweight and obese pregnant women. *Sci. Rep*. 2017; 7: 2860. DOI: 10.1038/s41598-017-03066-4.
194. Mackie R.I., Sghir A., Gaskins H.R. Developmental microbial ecology of the neonatal gastrointestinal tract. In *American Journal of Clinical Nutrition*. 1999; 69(5): 1035S-1045S. June 1999 with 629 Reads.
195. Hesla H.M., Stenius F., Jäderlund L. et al. Impact of lifestyle on the gut microbiota of healthy infants and their mothers — the ALADDIN birth cohort. *Microbiol Ecol*. 2014; 90 (3): 791–801.
196. Yudina Yu.V., Aminova A.I., Prodeus A.P. i dr. Osobennosti mikrobioty kishechnika u detey v vozraste 1–5 let s atopicheskim dermatitom. [Features of the intestinal microbiota in children aged 1–5 years with atopic dermatitis]. *Voprosy detskoy diyetologii*. 2021; 19(2): 5–13. (in Russian).
197. Jakobsson H.E., Abrahamsson T.R., Jenmalm M.C. et al. Decreased gut microbiota diversity, delayed bacteroidetes colonisation and reduced th1 responses in infants delivered by caesarean section. *Gut*. 2014; 63(4): 559–66.
198. MacIntyre D.A., Chandiramani M., Lee Y.S. et al. The vaginal microbiome during pregnancy and the postpartum period in a European population. *Sci Rep*. 2015; 5: 8988.
199. Jost T., Lacroix C., Braegger C., Chassard C. Assessment of bacterial diversity in breast milk using culture-dependent and culture-independent approaches. *Br J Nutr*. 2013; 110(7): 1253–62.
200. Soto A., Martín V., Jiménez E. et al. Lactobacilli and bifidobacteria in human breast milk: influence of antibiotherapy and other host and clinical factors. *J Pediatr Gastroenterol Nutr*. 2014; 59(1): 78–88.
201. Zivkovic A.M., German J.B., Lebrilla C.B., Mills D.A. Human milk glycobiome and its impact on the infant gastrointestinal microbiota. *Proc Natl Acad Sci USA*. 2011; 108(1): 4653–61.
202. Garrido D., Ruiz-Moyano S., Mills D.A. Release and utilization of N-acetyl-D-glucosamine from human milk oligosaccharides by *Bifidobacterium longum* subsp. *Infantis Anaerobe*. 2012; 18(4): 430–35.
203. Penders J., Thijs C., Vink C. et al. Factors influencing the composition of the intestinal microbiota in early infancy. *Pediatrics*. 2006; 118(2): 511–21.
204. Rutayisire E., Huang K., Liu Y., Tao F. The mode of delivery affects the diversity and colonization pattern of the gut microbiota during the first year of infants' life: a systematic review. *BMC Gastroenterology*. 2016; 16: 86.
205. Panda S., El khader I., Casellas F. et al. Short-term effect of antibiotics on human gut microbiota. *PLoS One*. 2014; 9(4): e95476.
206. Sonnenburg J.L., Sonnenburg E.D. Vulnerability of the industrialized microbiota. *Science*. 2019; 366: eaaw9255.
207. Stark C.M., Susi A., Emerick J., Nylund C.M. Antibiotic and acid-suppression medications during early childhood are associated with obesity. *Gut*. 2019; 68: 62–9.
208. Kronman M.P., Zaoutis T.E., Haynes K. et al. Antibiotic exposure and iBD development among children: A population-based cohort study. *Pediatrics* 2012; 130: e794–e803.
209. Langdon A., Crook N., Dantas G. The effects of antibiotics on the microbiome through out development and alternative approaches for therapeutic modulation. *Genome Med*. 2016; 8(1): 39.
210. Turnbaugh P.J., Ley R.E., Mahowald M.A. et al. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*. 2006; 444: 1027–31. DOI: 10.1038/nature05414.
211. Schwartz A., Taras D., Schäfer K. et al. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity*. 2010; 18: 190–5. DOI: 10.1038/oby.2009.167.
212. Turnbaugh P.J., Quince C., Faith J.J. et al. Organismal, genetic, and transcriptional variation in the deeply sequenced gut microbiomes of identical twins. *Proc. Natl. Acad. Sci. USA*. 2010; 107: 7503–8. DOI: 10.1073/pnas.1002355107.
213. Waldram A., Holmes E., Wang Y. et al. Top-down systems biology modeling of host metabolite-microbiome associations in obese rodents. *J. Proteome Res*. 2009; 8: 2361–75. DOI: 10.1021/pr8009885.
214. Zhang H., DiBaise J.K., Zuccolo A. et al. Human gut microbiota in obesity and after gastric bypass. *Proc. Natl. Acad. Sci. USA*. 2009; 106: 2365–70. DOI: 10.1073/pnas.0812600106.
215. Cuevas-Sierra A., Ramos-Lopez O., Riezu-Boj J.I. et al. Diet, Gut Microbiota, and Obesity: Links with Host Genetics and Epigenetics and Potential Applications. *Adv. Nutr*. 2019; 10: S17–S30. DOI: 10.1093/advances/nmy078.

216. Hiippala K., Jouhten H., Ronkainen A. et al. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients*. 2018; 10: 988. DOI: 10.3390/nu10080988.
217. Cani P.D., Amar J., Iglesias M.A. et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007; 56: 1761–72. DOI: 10.2337/db06-1491.
218. Xu P., Li M., Zhang J., Zhang T. Correlation of intestinal microbiota with overweight and obesity in Kazakh school children. *BMC Microbiol.* 2012; 12: 283. DOI: 10.1186/1471-2180-12-283.
219. Munukka E., Wiklund P., Pekkala S. et al. Women with and without metabolic disorder differ in their gut microbiota composition. *Obesity*. 2012; 20: 1082–7. DOI: 10.1038/oby.2012.8.
220. Kravtsova K., Prokopieva N.E., Petrenko Yu.V. et al. Gut microbiota of children born to obese mothers. *World of Microbiome*, Vienna, 28–30 апреля 2022 года. Vienna: Kenes group. 2022.
221. Gagliardi A., Totino V., Cacciotti F. et al. Rebuilding the Gut Microbiota Ecosystem. *Int. J. Environ. Res. Public Health*. 2018; 15: 1679. DOI: 10.3390/ijerph15081679.
222. Zhao Y., He X., Shi X. et al. Association between serum amyloid A and obesity: A meta-analysis and systematic review. *Inflamm. Res.* 2010; 59: 323–34. DOI: 10.1007/s00011-010-0163-y.
223. Payne A.N., Chassard C., Zimmermann M. et al. The metabolic activity of gut microbiota in obese children is increased compared with normal-weight children and exhibits more exhaustive substrate utilization. *Nutr. Diabetes*. 2011; 1: e12. DOI: 10.1038/nutd.2011.8.
224. Alex S., Lichtenstein L., Dijk W. et al. ANGPTL4 is produced by entero-endocrine cells in the human intestinal tract. *Histochem. Cell Biol.* 2014; 141: 383–91. DOI: 10.1007/s00418-013-1157-y.
225. Sanmiguél C., Gupta A., Mayer E.A. Gut Microbiome and Obesity: A Plausible Explanation for Obesity. *Curr. Obes. Rep.* 2015; 4: 250–61. DOI: 10.1007/s13679-015-0152-0.
226. Stephens R.W., Arhire L., Covasa M. Gut Microbiota: From Microorganisms to Metabolic Organ Influencing Obesity. *Obesity*. 2018; 26: 801–9. DOI: 10.1002/oby.22179.
227. Bäckhed F., Ding H., Wang T. et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. USA*. 2004; 101: 15718–23. DOI: 10.1073/pnas.0407076101.
228. Scarpellini E., Cazzato A., Lauritano C. et al. Probiotics: Which and when? *Dig. Dis.* 2008; 26: 175–82. DOI: 10.1159/000116776.
229. Smirnova N.N., Novikova V.P., Kupriyenko N.B. i dr. Vliyaniye mikrobioma reproduktivnogo trakta zhenshchiny na vnutritrobnoye i postnatal'noye razvitiye rebenka. [Influence of the female reproductive tract microbiome on intrauterine and postnatal development of the child]. *Voprosy ginekologii, akusherstva i perinatologii*. 2022; 21(6): 107–13. DOI: 10.20953/1726-1678-2022-6-107-112. EDN LCUFFM. (in Russian).
230. Duncan S.H., Belenguer A., Holtrop G. Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate-producing bacteria in feces. *Appl. Environ. Microbiol.* 2007; 73: 1073–8. DOI: 10.1128/AEM.02340-06.
231. Murphy E.F., Cotter P.D., Healy S. et al. Composition and energy harvesting capacity of the gut microbiota: Relationship to diet, obesity and time in mouse models. *Gut*. 2010; 59: 1635–42. DOI: 10.1136/gut.2010.215665.
232. Li J.V., Ashrafi H., Bueter M. et al. Metabolic surgery profoundly influences gut microbial-host metabolic cross-talk. *Gut*. 2011; 60: 1214–23. DOI: 10.1136/gut.2010.234708.
233. Liou A.P., Paziuk M., Luevano J.M., Jr. et al. Conserved shifts in the gut microbiota due to gastric bypass reduce host weight and adiposity. *Sci. Transl. Med.* 2013; 5: 178ra41. DOI: 10.1126/scitranslmed.3005687.
234. Hiippala K., Jouhten H., Ronkainen A. et al. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients*. 2018; 10: 988. DOI: 10.3390/nu10080988.
235. Cani P.D., Amar J., Iglesias M.A. et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007; 56: 1761–72. DOI: 10.2337/db06-1491.
236. Lee S., Sung J., Lee J., Ko G. Comparison of the gut microbiotas of healthy adult twins living in South Korea and the United States. *Appl. Environ. Microbiol.* 2011; 77: 7433–7. DOI: 10.1128/AEM.05490-11.
237. Luoto R., Kalliomäki M., Laitinen K. et al. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J. Pediatr. Gastroenterol. Nutr.* 2011; 52: 90–5. DOI: 10.1097/MPG.0b013e-3181f3457f.
238. Payne A.N., Chassard C., Banz Y., Lacroix C. The composition and metabolic activity of child gut microbiota demonstrate differential adaptation to varied nutrient loads in an in vitro model of colonic fermentation. *FEMS Microbiol. Ecol.* 2012; 80: 608–23. DOI: 10.1111/j.1574-6941.2012.01330.x.
239. Gohir W., Whelan F.J., Surette M.G. et al. Pregnancy-related changes in the maternal gut microbiota are dependent upon the mother's periconcep-

- tional diet. *Gut Microbes*. 2015; 6: 310–20. DOI: 10.1080/19490976.2015.1086056.
240. Collado M.C., Isolauri E., Laitinen K. and Salminen S. Distinct composition of gut microbiota during pregnancy in overweight and normal-weight women. *Am. J. Clin. Nutr.* 2008; 88: 894–9.
 241. Qin J., Li R., Raes J. et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature*. 2010; 464: 59–65. DOI: 10.1038/nature08821.
 242. Dietert R.R. and Dietert J.M. The microbiome and sustainable healthcare. *Healthcare (Basel)*. 2015; 3: 100–29. DOI: 10.3390/healthcare3010100.
 243. Zacarías M.F., Collado M.C., Gómez-Gallego C. et al. Pregestational overweight and obesity are associated with differences in gut microbiota composition and systemic inflammation in the third trimester. *PLoS One*. 2018; 13(7): e0200305. DOI: 10.1371/journal.pone.0200305. PMID: 30005082; PMCID: PMC6044541.
 244. Santacruz A., Collado M.C., Garcia-Valdes L. et al. Gut microbiota composition is associated with body weight, weight gain and biochemical parameters in pregnant women. *Br. J. Nutr.* 2010; 104: 83–92. DOI: 10.1017/S0007114510000176.
 245. Soderborg T.K., Clark S.E., Mulligan C.E. et al. The gut microbiota in infants of obese mothers increases inflammation and susceptibility. *Nat. Commun.* 2018; 9(1). DOI: 10.1038/s41467-018-06929-0.
 246. Ferrer M., Ruiz A., Lanza F. et al. Microbiota from the distal guts of lean and obese adolescents exhibit partial functional redundancy besides clear differences in community structure. *Environ. Microbiol.* 2013; 15: 211–26. DOI: 10.1111/j.1462-2920.2012.02845.x.
 247. Bervoets L., Van Hoorenbeeck K., Kortleven I. et al. Differences in gut microbiota composition between obese and lean children: A cross-sectional study. *Gut Pathog.* 2013; 5: 10. DOI: 10.1186/1757-4749-5-10.
 248. Luoto R., Kalliomäki M., Laitinen K. et al. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J. Pediatr. Gastroenterol. Nutr.* 2011; 52: 90–5. DOI: 10.1097/MPG.0b013e-3181f3457f.
 249. Clarke S.F., Murphy E.F., O'Sullivan O. et al. Targeting the microbiota to address diet-induced obesity: A time dependent challenge. *PLoS One*. 2013; 8: e65790. DOI: 10.1371/journal.pone.0065790.
 250. Prokop'yeva N. E., Petrenko Yu.V., Ivanov D.O. i dr. Formirovaniye polostnoy mikrobioty kishcheznika u detey pervogo goda zhizni, rozhdennykh ot materey s ozhireniyem. [Formation of cavity intestinal microbiota in children of the first year of life born to obese mothers]. Aktual'nyye problemye abdominal'noy patologii u detey: Materialy Yubileynogo XXX Kongressa detskikh gastroenterologov Rossii i stran SNG, Moskva, 14–16 marta 2023 goda. Moskva: Medpraktika-M Publ.; 2023: 41–3. (in Russian).
 251. Martinez I., Lattimer J.M., Hubach K.L. et al. Gut microbiome composition is linked to whole grain-induced immunological improvements. *Isme J.* 2013; 7: 269–80. DOI: 10.1038/ismej.2012.104.
 252. Vael C., Verhulst S.L., Nelen V. et al. Intestinal microbiota and body mass index during the first three years of life: An observational study. *Gut Pathog.* 2011; 3: 8. DOI: 10.1186/1757-4749-3-8.
 253. Nadal I., Santacruz A., Marcos A. et al. Shifts in clostridia, bacteroides and immunoglobulin-coating fecal bacteria associated with weight loss in obese adolescents. *Int. J. Obes.* 2009; 33: 758–67. DOI: 10.1038/ijo.2008.260.

ЛИТЕРАТУРА

1. World Health Organization (WHO). Obesity and overweight. January 2015. <http://www.who.int/mediacentre/factsheets/fs311/en/>. Accessed 2 April 2016.
2. Hruby A., Hu F.B. The epidemiology of obesity: a big picture. *Pharmacoeconomics*. 2015; 33: 673–89.
3. Poston L., Caleyachetty R., Cnattingius S. et al. Pre-conceptional and maternal obesity: epidemiology and health consequences. *Lancet Diabetes Endocrinol.* 2016; 4(12): 1025–36. DOI: 10.1016/S2213-8587(16)30217-0.
4. Hales C.M., Carroll M.D., Fryar C.D., Ogden C.L. Prevalence of Obesity and Severe Obesity Among Adults: United State-2018. *NCHS Data Brief*. 2020; 360: 1–8.
5. Грицинская В.Л., Новикова В.П., Хавкин А.И. К вопросу об эпидемиологии ожирения у детей и подростков (систематический обзор и мета-анализ научных публикаций за 15-летний период). *Вопросы практической педиатрии*. 2022; 17(2): 126–35. DOI: 10.20953/1817-7646-2022-2-126-135.
6. Gaillard R., Steegers EAP, Franco O.H. et al. Maternal weight gain in different periods of pregnancy and childhood cardio-metabolic outcomes. The Generation R Study. *International Journal of Obesity*. 2015; 39: 677–85.
7. Hinkle S.N., Sharma A.J., Swan D.W. et al. Excess Gestational Weight Gain Is Associated with Child Adiposity among Mothers with Normal and Overweight Prepregnancy Weight Status. *The Journal of Nutrition*. 2012; 142: 1851–8.
8. Hochner H., Friedlander Y., Calderon-Margalit R. et al. Associations of maternal prepregnancy body

- mass index and gestational weight gain with adult offspring cardiometabolic risk factors: the Jerusalem Perinatal Family Follow-up Study. *Circulation*. 2012; 125: 1381–9.
9. Oken E., Rifas-Shiman S.L., Field A.E. et al. Maternal Gestational Weight Gain and Offspring Weight in Adolescence. 2008; 112: 8.
 10. Kaar J.L., Crume T., Brinton J.T. et al. Maternal Obesity, Gestational Weight Gain, and Offspring Adiposity: The Exploring Perinatal Outcomes among Children Study. *The Journal of Pediatrics*. 2014; 165: 509–15.
 11. Josey M.J., McCullough L.E., Hoyo C., Williams-DeVane C. Overall gestational weight gain mediates the relationship between maternal and child obesity. *BMC public health*. 2019; 19: 1062.
 12. Arrowsmith S., Wray S., Quenby S. Maternal obesity and labour complications following induction of labour in prolonged pregnancy. *BJOG*. 2011; 118(5): 578–88.
 13. Yogev Y., Catalano P.M. Pregnancy and obesity. *Obstet. Gynecol. Clin. North Am.* 2009; 36(2): 285–300.
 14. Bogaerts A., Witters I., Van den Bergh B.R. et al. Obesity in pregnancy: altered onset and progression of labour. *Midwifery*. 2013; 29(12): 1303–13.
 15. HAPO Study Cooperative Research Group, Metzger B.E., Lowe L.P., Dyer A.R., Trimble E.R., Chaovarindr U. et al. Hyperglycemia and adverse pregnancy outcomes. *N Engl J Med*. 2008; 358(19): 1991–2002.
 16. Wu D., Hu D., Chen H. et al. Glucose-regulated phosphorylation of TET2 by AMPK reveals a pathway linking diabetes to cancer. *Nature*. 2018; 559: 637–41.
 17. Комшилова К.А., Дзгоева Ф.Х. Беременность и ожирение. *Ожирение и метаболизм*. 2009; 4: 9–13.
 18. Чухарева Н.А., Рунихина Н.К., Дудинская Е.Н. Особенности течения беременности у женщин с ожирением. *Акушерство и гинекология*. 2014; 2: 9–13.
 19. Lazo-de-la-Vega-Monroy M-L., Mata-Tapia K-A., Garcia-Santillan J-A. et al. Association of placental nutrient sensing pathways with birth weight. *Reproduction*. 2020; 160: 455–68.
 20. Zhu J. T Helper Cell Differentiation, Heterogeneity, and Plasticity. *Cold Spring Harb. Perspect. Biol.* 10(10), 1–17. DOI: 10.1101/cshperspect.a030338.
 21. Zhu M.J., Du M., Nathanielsz P.W., Ford S.P. Maternal obesity up-regulates inflammatory signaling pathways and enhances cytokine expression in the mid-gestation sheep placenta. *Placenta*. 2010; 31(5): 387–91. DOI: 10.1016/j.placenta.2010.02.002.
 22. Romagnani S. T-cell subsets (Th1 versus Th2). *Ann. Allergy Asthma Immunol.* 2000; 85(1): 9–18. DOI: 10.1016/S1081-1206(10)62426-X.
 23. Liang T., Jinglong X., Shusheng D., Aiyu W. Maternal obesity stimulates lipotoxicity and up-regulates inflammatory signaling pathways in the full-term swine placenta. *Anim Sci J*. 2018; 89: 1310–22.
 24. Brass E., Hanson E., O'Tierney-Ginn P.F. Placental oleic acid uptake is lower in male offspring of obese women. *Placenta*. 2013; 34: 503–9.
 25. Jansson N., Rosario F.J., Gaccioli F. et al. Activation of placental mTOR signaling and amino acid transporters in obese women giving birth to large babies. *J Clin Endocrinol Metab*. 2013; 98: 105–13.
 26. Flenady V., Koopmans L., Middleton P. et al. Major risk factors for stillbirth in high-income countries: a systematic review and meta-analysis. *Lancet Lond Engl*. 2011; 377(9774): 1331–40.
 27. Aune D., Saugstad O.D., Henriksen T., Tonstad S. Maternal body mass index and the risk of fetal death, stillbirth, and infant death: a systematic review and meta-analysis. *JAMA*. 2014; 311: 1536–46.
 28. Ehrenberg H.M., Mercer B.M., Catalano P.M. The influence of obesity and diabetes on the prevalence of macrosomia. *Am J Obstet Gynecol*. 2004; 191: 964–8.
 29. Sewell M.F., Huston-Presley L., Super D.M. et al. Increased neonatal fat mass, not lean body mass, is associated with maternal obesity. *Am J Obstet Gynecol*. 2006; 195: 1100–3.
 30. Whitelaw A.G. Influence of maternal obesity on subcutaneous fat in the newborn. *Br Med J*. 1976; 1: 985–6.
 31. Andres A., Hull H.R., Shankar K. et al. Longitudinal body composition of children born to mothers with normal weight, overweight, and obesity. *Obes Silver Spring Md*. 2015; 23: 1252–8.
 32. Jolly M.C., Sebire N.J., Harris J.P. et al. Risk factors for macrosomia and its clinical consequences: a study of 350, 311 pregnancies. *Eur J Obstet Gynecol Reprod Biol*. 2003; 111(1): 9–14.
 33. Nesbitt T.S., Gilbert W.M., Herrchen B. Shoulder dystocia and associated risk factors with macrosomic infants born in California. *Am J Obstet Gynecol*. 1998; 179(2): 476–80.
 34. Turner D., Monthé-Drèze C., Cherkerzian S. et al. Maternal obesity and cesarean section delivery: additional risk factors for neonatal hypoglycemia? *J Perinatol Off J Calif Perinat Assoc*. 2019; 39(8): 1057–64.
 35. Stanley C.A., Rozance P.J., Thornton P.S. et al. Re-evaluating “transitional neonatal hypoglycemia”: mechanism and implications for management. *J Pediatr*. 2015; 166(6): 1520–5.e1.

36. Gimeno R.E., Klamann L.D. Adipose tissue as an active endocrine organ: recent advances. 2005; 5: 122–8.
37. Hutley L., Prins J.B. Fat as an endocrine organ: relationship to the metabolic syndrome. 2005; 330: 280–9.
38. Kahn B.B., Flier J.S. Obesity and insulin resistance. 2000; 106: 473–81.
39. Kahn S.E., Hull R.L., Utzschneider K.M. Mechanisms linking obesity to insulin resistance and type 2 diabetes. 2006; 444: 840–6.
40. Matsuzawa Y., Funahashi T., Nakamura T. Molecular mechanism of metabolic syndrome X: contribution of adipocytokines adipocyte-derived bioactive substances. 1999; 892: 146–54.
41. Montague C.T., O'Rahilly S. The perils of portliness: causes and consequences of visceral adiposity. 2000; 49: 883–8.
42. Ronti T., Lupattelli G., Mannarino E. The endocrine function of adipose tissue: an update. 2006; 64: 355–65.
43. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. 2001; 104: 531–43.
44. Trayhurn P. Endocrine and signalling role of adipose tissue: new perspectives on fat. 2005; 184: 285–93.
45. Catalano P.M., Hoegh M., Minium J. et al. Adiponectin in human pregnancy: implications for regulation of glucose and lipid metabolism. 2006; 49: 1677–85.
46. Farvid M.S., Ng T.W., Chan D.C. et al. Association of adiponectin and resistin with adipose tissue compartments, insulin resistance and dyslipidaemia. 2005; 7: 406–13.
47. Gable D.R., Hurel S.J., Humphries S.E. Adiponectin and its gene variants as risk factors for insulin resistance, the metabolic syndrome and cardiovascular disease. 2006; 188: 231–44.
48. Kirwan J.P., Hauguel-De M.S., Lepercq J. et al. TNF-alpha is a predictor of insulin resistance in human pregnancy. 2002; 51: 2207–13.
49. Lopez-Bermejo A., Fernandez-Real J.M., Garrido E. et al. Maternal soluble tumour necrosis factor receptor type 2 (sTNFR2) and adiponectin are both related to blood pressure during gestation and infant's birthweight. 2004; 61: 544–52.
50. Matsuzawa Y. The metabolic syndrome and adipocytokines. 2006; 580: 2917–21.
51. McLachlan K.A., O'Neal D., Jenkins A., Alford F.P. Do adiponectin, TNFalpha, leptin and CRP relate to insulin resistance in pregnancy? Studies in women with and without gestational diabetes, during and after pregnancy. *Diabetes Metab Res Rev.* 2006; 22: 131–8.
52. Ouchi N., Kihara S., Arita Y. et al. Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF-kappaB signaling through a cAMP-dependent pathway. 2000; 102: 1296–1301.
53. Retnakaran R., Hanley A.J., Raif N. et al. Reduced adiponectin concentration in women with gestational diabetes: a potential factor in progression to type 2 diabetes. 2004; 27: 799–800.
54. Silha J.V., Krsek M., Skrha J.V. et al. Plasma resistin, adiponectin and leptin levels in lean and obese subjects: correlations with insulin resistance. 2003; 149: 331–5.
55. Steppan C.M., Lazar M.A. Resistin and obesity-associated insulin resistance. 2002; 13: 18–23.
56. Tilg H., Moschen A.R. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. 2006; 6: 772–83.
57. Unger R.H. Hyperleptinemia: protecting the heart from lipid overload. 2005; 45: 1031–4.
58. Смирнова Н.Н., Куприенко Н.Б., Петренко Ю.В., Новикова В.П. Материнское ожирение и система «мать-плацента-плод»: доказанные механизмы влияния. *Children's Medicine of the North-West.* 2021; 9(3): 31–9.
59. Yokota T., Oritani K., Takahashi I. et al. Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the functions of macrophages. 2000; 96: 1723–32.
60. Harmon K.A., Gerard L., Jensen D.R. et al. Continuous glucose profiles in obese and normal-weight pregnant women on a controlled diet: metabolic determinants of fetal growth. *Diabetes Care.* 2011; 34(10): 2198–204. DOI: 10.2337/dc11-0723.
61. Смирнова Н.Н., Хавкин А.И., Новикова В.П. Состав грудного молока при ожирении матери: влияние на развитие ребенка. *Вопросы практической педиатрии.* 2022; 17(1): 167–76. DOI: 10.20953/1817-7646-2022-1-167-176.
62. Цепилова М.О., Полякова К.Д. Влияние активных метаболитов грудного молока и их производных на организм новорожденного. *Проба пера: Материалы межрегиональной научной конференции молодых ученых «VI Малые Апрельские чтения памяти профессора М.В. Пиккель»*, Архангельск, 01 апреля 2023 года. Выпуск 6. Архангельск: Северный государственный медицинский университет; 2023.
63. Смирнова Н.Н., Хавкин А.И., Куприенко Н.Б., Новикова В.П. Бактерии и вирусы грудного молока. *Вопросы детской диетологии.* 2022; 20(2): 74–82. DOI: 10.20953/1727-5784-2022-2-74-82. EDN BBIKOO.
64. Metzger B.E., Lowe L.P., Dyer A.R. et al. Hyperglycemia and adverse pregnancy outcomes. *N Engl J*

- Med. 2008; 358(19): 1991–2002. DOI: 10.1056/NEJ-Moa0707943.
65. Xiang A.H., Peters R.K., Trigo E. et al. Multiple metabolic defects during late pregnancy in women at high risk for type 2 diabetes. *Diabetes*. 1999; 48(4): 848–54. DOI: 10.2337/diabetes.48.4.848.
66. Friedman J.E., Ishizuka T., Shao J. et al. Impaired glucose transport and insulin receptor tyrosine phosphorylation in skeletal muscle from obese women with gestational diabetes. *Diabetes*. 1999; 48(9): 1807–14. DOI: 10.2337/diabetes.48.9.1807.
67. Catalano P.M., Ehrenberg H.M. The short- and long-term implications of maternal obesity on the mother and her offspring. *BJOG*. 2006; 113(10): 1126–33. DOI: 10.1111/j.1471-0528.2006.00989.x.
68. Catalano P.M., Presley L., Minium J., Hauguel-de Mouzon S. Fetuses of obese mothers develop insulin resistance in utero. *Diabetes Care*. 2009; 32(6): 1076–80. DOI: 10.2337/dc08-2077.
69. Toran-Allerand C.D., Ellis L., Pfenninger K.H. Estrogen and insulin synergism in neurite growth enhancement in vitro: mediation of steroid effects by interactions with growth factors? *Brain Res*. 1988; 469(1-2): 87–100. DOI: 10.1016/0165-3806(88)90172-1.
70. Recio-Pinto E., Ishii D.N. Effects of insulin, insulin-like growth factor-II and nerve growth factor on neurite outgrowth in cultured human neuroblastoma cells. *Brain Res*. 1984; 302(2): 323–34. DOI: 10.1016/0006-8993(84)90246-4.
71. Lázár B.A., Jancsó G., Pálvolgyi L. et al. Insulin confers differing effects on neurite outgrowth in separate populations of cultured dorsal root ganglion neurons: The role of the insulin receptor. *Front Neurosci*. 2018; 12: 732. DOI: 10.3389/fnins.2018.00732.
72. Song J., Wu L., Chen Z. et al. Axons guided by insulin receptor in drosophila visual system. *Science*. 2003; 300(5618): 502–5. DOI: 10.1126/science.1081203.
73. Fex Svenningsen A., Kanje M. Insulin and the insulin-like growth factors I and II are mitogenic to cultured rat sciatic nerve segments and stimulate [3H]thymidine incorporation through their respective receptors. *Glia*. 1996; 18(1): 68–72. DOI: 10.1002/(SICI)1098-1136(199609)18:1.
74. Dudek H., Datta S.R., Franke T.F. et al. Regulation of neuronal survival by the serine-threonine protein kinase akt. *Science*. 1997; 275(5300): 661–5. DOI: 10.1126/science.275.5300.661.
75. Apostolatos A., Song S., Acosta S. et al. Insulin promotes neuronal survival via the alternatively spliced protein kinase C δ II isoform. *J Biol Chem*. 2012; 287(12): 9299–310. DOI: 10.1074/jbc.M111.313080.
76. Haddad-Tóvolli R., Altirriba J., Obri A. et al. Pro-opiomelanocortin (POMC) neuron translational signatures underlying obesogenic gestational malprogramming in mice. *Mol Metab*. 2020; 36: 100963. DOI: 10.1016/j.molmet.2020.02.006.
77. Melo A.M., Benatti R.O., Ignacio-Souza L.M. et al. Hypothalamic endoplasmic reticulum stress and insulin resistance in offspring of mice dams fed high-fat diet during pregnancy and lactation. *Metabolism*. 2014; 63(5): 682–92. DOI: 10.1016/j.metabol.2014.02.002.
78. Wang Z.V., Scherer P.E. Adiponectin, the past two decades. *J Mol Cell Biol*. 2016; 8: 93–100.
79. Chandran M., Phillips S.A., Ciaraldi T., Henry R.R. Adiponectin: more than just another fat cell hormone? *Diabetes Care*. 2003; 26: 2442–50.
80. Arita Y., Kihara S., Ouchi N. et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. 1999; 257: 79–83.
81. Hu E., Liang P., Spiegelman B.M. AdipoQ is a novel adipose-specific gene dysregulated in obesity. 1996; 271: 10697–703.
82. Hinkle S.N., Rawal S., Liu D. et al. Maternal adipokines longitudinally measured across pregnancy and their associations with neonatal size, length, and adiposity. *Int J Obes (Lond)*. 2019; 43: 1422–34.
83. Aye I.L., Powell T.L., Jansson T. Review: Adiponectin — the missing link between maternal adiposity, placental transport and fetal growth? *Placenta*. 2013; 34: S40–5.
84. Qiao L., Yoo H.S., Madon A. et al. Adiponectin enhances mouse fetal fat deposition. *Diabetes*. 2012; 61: 3199–207.
85. Forhead A.J., Fowden A.L. The hungry fetus? Role of leptin as a nutritional signal before birth. *J Physiol*. 2009; 587: 1145–52.
86. Pardo I.M., Geloneze B., Tambascia M.A., Barros-Filho A.A. Hyperadiponectinemia in newborns: relationship with leptin levels and birth weight. *Obes Res*. 2004; 12: 521–4.
87. Frederiksen L., Nielsen T.L., Wraae K. et al. Subcutaneous rather than visceral adipose tissue is associated with adiponectin levels and insulin resistance in young men. *J Clin Endocrinol Metab*. 2009; 94: 4010–5.
88. Misra V.K., Straughen J.K., Trudeau S. Maternal serum leptin during pregnancy and infant birth weight: the influence of maternal overweight and obesity. *Obes (Silver Spring)*. 2013; 21(5): 1064–9. DOI: 10.1002/oby.20128.
89. Patro-Małyśza J., Trojnar M., Skórzyńska-Dziduszko K.E. et al. Leptin and ghrelin in excessive gestational weight gain-association between mothers and offspring. *Int J Mol Sci*. 2019; 20(10): 2398. DOI: 10.3390/ijms20102398.

90. Trayhurn P., Beattie J.H. Physiological role of adipose tissue: white adipose tissue as an endocrine and secretory organ. *Proc Nutr Soc.* 2001; 60: 329–39.
91. Morrison C.D. Leptin resistance and the response to positive energy balance. *Physiol Behav.* 2008; 94: 660–3.
92. Tham E., Liu J., Innis S. et al. Acylated ghrelin concentrations are markedly decreased during pregnancy in mothers with and without gestational diabetes: relationship with cholinesterase. *Am J Physiol Endocrinol Metab.* 2009; 296(5): E1093–100. DOI: 10.1152/ajpendo.90866.2008.
93. Karakulak M., Saygili U., Temur M. et al. Comparison of umbilical cord ghrelin concentrations in full-term pregnant women with or without gestational diabetes. *Endocr Res.* 2017; 42(2): 79–85. DOI: 10.1080/07435800.2016.1194855.
94. Nakahara K., Nakagawa M., Baba Y. et al. Maternal ghrelin plays an important role in rat fetal development during pregnancy. *Endocrinology.* 2006; 147(3): 1333–42. DOI: 10.1210/en.2005-0708.
95. Wang Y., Sul H.S. Ectodomain shedding of preadipocyte factor 1 (Pref-1) by tumor necrosis factor alpha converting enzyme (TACE) and inhibition of adipocyte differentiation. *Mol Cell Biol.* 2006; 26: 5421–35.
96. Shulman G.I. Cellular mechanisms of insulin resistance. *J Clin Invest.* 2000; 106: 171–6.
97. Garg A. Acquired and inherited lipodystrophies. *N Engl J Med.* 2004; 350: 1220–34.
98. Shackleton S., Lloyd D.J., Jackson S.N. et al. LMNA, encoding lamin A/C, is mutated in partial lipodystrophy. *Nat Genet.* 2000; 24: 153–6.
99. Agarwal A.K., Arioglu E., De Almeida S. et al. AGPAT2 is mutated in congenital generalized lipodystrophy linked to chromosome 9q34. *Nat Genet.* 2002; 31: 21–3.
100. Magre J., Delepine M., Khallouf E. et al. Identification of the gene altered in Berardinelli-Seip congenital lipodystrophy on chromosome 11q13. *Nat Genet.* 2001; 28: 365–70.
101. Barroso I., Gurnell M., Crowley V.E. et al. Dominant negative mutations in human PPARgamma associated with severe insulin resistance, diabetes mellitus and hypertension. *Nature.* 1999; 402: 880–3.
102. Gregoire F.M., Smas C.M., Sul H.S. Understanding adipocyte differentiation. *Physiol Rev.* 1998; 78: 783–809.
103. Rosen E.D., Spiegelman B.M. Molecular regulation of adipogenesis. *Annu Rev Cell Dev Biol.* 2000; 16: 145–71.
104. Smas C.M., Sul H.S. Pref-1, a protein containing EGF-like repeats, inhibits adipocyte differentiation. *Cell.* 1993; 73: 725–34.
105. Bonen A., Parolin M.L., Steinberg G.R. et al. Triacylglycerol accumulation in human obesity and type 2 diabetes is associated with increased rates of skeletal muscle fatty acid transport and increased sarcolemmal FAT/CD36. *Faseb J.* 2004; 18: 1144–6.
106. Smas C.M., Chen L., Sul H.S. Cleavage of membrane-associated pref-1 generates a soluble inhibitor of adipocyte differentiation. *Mol Cell Biol.* 1997; 17: 977–88.
107. Kershaw E.E., Flier J.S. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab.* 2004; 89: 2548–56.
108. Villena J.A., Kim K.H., Sul H.S. Pref-1 and ADSF/resistin: two secreted factors inhibiting adipose tissue development. *Horm Metab Res.* 2002; 34: 664–70.
109. Furigo I.C., Teixeira P.D.S., de Souza G.O. et al. Growth hormone regulates neuroendocrine responses to weight loss via AgRP neurons. *Nat Commun.* 2019; 10(1): 662. DOI: 10.1038/s41467-019-08607-1.
110. Furigo I.C., de Souza G.O., Teixeira P.D.S. et al. Growth hormone enhances the recovery of hypoglycemia. *FASEB J.* 2019; 33(11): 11909–24. DOI: 10.1096/fj.201901315R.
111. Donato J., Wasinski F., Furigo I.C. et al. Central regulation of metabolism by growth hormone. *Cells.* 2021; 10(1): 129. DOI: 10.3390/cells10010129.
112. Teixeira P.D.S., Couto G.C., Furigo I.C. et al. Central growth hormone action regulates metabolism during pregnancy. *Am J Physiol Endocrinol Metab.* 2019; 317(5): E925–E40. DOI: 10.1152/ajpendo.00229.2019.
113. Wasinski F., Furigo I.C., Teixeira P.D.S. et al. Growth hormone receptor deletion reduces the density of axonal projections from hypothalamic arcuate nucleus neurons. *Neuroscience.* 2020; 434: 136–47. DOI: 10.1016/j.neuroscience.2020.03.037.
114. Challier J.C., Basu S., Bintein T. et al. Obesity in pregnancy stimulates macrophage accumulation and inflammation in the placenta. *Placenta.* 2008; 29(3): 274–81.
115. Ramsay J.E., Ferrell W.R., Crawford L. et al. Maternal obesity is associated with dysregulation of metabolic, vascular, and inflammatory pathways. *J Clin Endocrinol Metab.* 2002; 87(9): 4231–7.
116. Петренко Ю.В., Герасимова К.С., Новикова В.П. Биологическая и патофизиологическая значимость адипонектина. *Педиатр.* 2019; 10(2): 83–7. DOI: 10.17816/PED10283-87. EDN RVWXCN.
117. Novikova V.P., Ivanov D.O., Petrenko Yu.V. et al. Increased marker of endothelial cell dysfunction sVCAM-1 in umbilical cord blood in neonates born to obese women. *Archives of Disease in*

- Childhood. 2019; 104(S3): 117. DOI 10.1136/archdis-child-2019-epa.273.
118. Azizian M., Mahdipour E., Mirhafez S.R. et al. Cytokine profiles in overweight and obese subjects and normal weight individuals matched for age and gender. *Ann Clin Biochem.* 2016; 53(6): 663–8.
119. Brunner S., Schmid D., Hüttinger K. et al. Maternal insulin resistance, triglycerides and cord blood insulin in relation to post-natal weight trajectories and body composition in the offspring up to 2 years. *Diabet Med J Br Diabet Assoc.* 2013; 30(12): 1500–7.
120. Regnault N., Botton J., Heude B. et al. Higher Cord C-Peptide Concentrations Are Associated With Slower Growth Rate in the 1st Year of Life in Girls but Not in Boys. *Diabetes.* 2011; 60(8): 2152–9.
121. Stang J., Huffman L.G. Position of the Academy of Nutrition and Dietetics: Obesity, Reproduction, and Pregnancy Outcomes. *J Acad Nutr Diet.* 2016; 116(4): 677–91.
122. Dubé E., Gravel A., Martin C. et al. Modulation of fatty acid transport and metabolism by maternal obesity in the human full-term placenta. *Biol Reprod.* 2012; 87(1): 14, 1–11.
123. Hellmuth C., Lindsay K.L., Uhl O. et al. Association of maternal prepregnancy BMI with metabolomic profile across gestation. *Int J Obes* 2005. 2017; 41(1): 159–69.
124. Briley A.L., Barr S., Badger S. et al. A complex intervention to improve pregnancy outcome in obese women; the UPBEAT randomised controlled trial. *BMC Pregnancy Childbirth.* 2014; 14: 74.
125. Herrera E., Ortega-Senovilla H. Implications of lipids in neonatal body weight and fat mass in gestational diabetic mothers and non-diabetic controls. *Curr Diabetes Rep.* 2018; 18(2): 7. DOI: 10.1007/s11892-018-0978-4.
126. Merzouk H., Meghelli-Bouchenak M., Loukidi B. et al. Impaired serum lipids and lipoproteins in fetal macrosomia related to maternal obesity. *Biol Neonate.* 2000; 77(1): 17–24. DOI: 10.1159/000014190.
127. Furse S., Koulman A., Ozanne S.E. et al. Altered lipid metabolism in obese women with gestational diabetes and associations with offspring adiposity. *J Clin Endocrinol Metab.* 2022; 107(7): e2825–e32. DOI: 10.1210/clinem/dgac206.
128. Barbour L.A., Farabi S.S., Friedman J.E. et al. Postprandial triglycerides predict newborn fat more strongly than glucose in women with obesity in early pregnancy. *Obes (Silver Spring).* 2018; 26(8): 1347–56. DOI: 10.1002/oby.22246.
129. Pimentel G.D., Lira F.S., Rosa J.C. et al. Intake of trans fatty acids during gestation and lactation leads to hypothalamic inflammation *via* TLR4/NFkBp65 signaling in adult offspring. *J Nutr Biochem.* 2012; 23(3): 265–71. DOI: 10.1016/j.jnutbio.2010.12.003.
130. Rother E., Kuschewski R., Alcazar M.A. et al. Hypothalamic JNK1 and IKK β activation and impaired early postnatal glucose metabolism after maternal perinatal high-fat feeding. *Endocrinology.* 2012; 153(2): 770–81. DOI: 10.1210/en.2011-1589.
131. Zhang X., Zhang G., Zhang H. et al. Hypothalamic IKK β /NF-kappaB and ER stress link overnutrition to energy imbalance and obesity. *Cell.* 2008; 135(1): 61–73. DOI: 10.1016/j.cell.2008.07.043.
132. Sadagurski M., Debarba L.K., Werneck-de-Castro J.P. et al. Sexual dimorphism in hypothalamic inflammation in the offspring of dams exposed to a diet rich in high fat and branched-chain amino acids. *Am J Physiol Endocrinol Metab.* 2019; 317(3): E526–E34. DOI: 10.1152/ajpendo.00183.2019.
133. Melo A.M., Benatti R.O., Ignacio-Souza L.M. et al. Hypothalamic endoplasmic reticulum stress and insulin resistance in offspring of mice dams fed high-fat diet during pregnancy and lactation. *Metabolism.* 2014; 63(5): 682–92. DOI: 10.1016/j.metabol.2014.02.002.
134. Park S., Jang A., Bouret S.G. Maternal obesity-induced endoplasmic reticulum stress causes metabolic alterations and abnormal hypothalamic development in the offspring. *PloS Biol.* 2020; 18(3): e3000296. DOI: 10.1371/journal.pbio.3000296.
135. Barnes S.K., Ozanne S.E. Pathways linking the early environment to long-term health and lifespan. *Prog Biophys Mol Biol.* 2011; 106(1): 323–36. DOI: 10.1016/j.pbiomolbio.2010.12.005.
136. Drake A.J., Reynolds R.M. Impact of maternal obesity on offspring obesity and cardiometabolic disease risk. *Reproduction.* 2010; 140(3): 387–98. DOI: 10.1530/REP-10-0077.
137. Gaillard R. Maternal obesity during pregnancy and cardiovascular development and disease in the offspring. *Eur J Epidemiol.* 2015; 30(11): 1141–52. DOI: 10.1007/s10654-015-0085-7.
138. Nicholas L.M., Morrison J.L., Rattanatrak L. et al. The early origins of obesity and insulin resistance: timing, programming and mechanisms. *Int J Obes (Lond).* 2016; 40(2): 229–38.
139. Poston L. Developmental programming and diabetes — the human experience and insight from animal models. *Best Pract Res Clin Endocrinol Metab.* 2010; 24(4): 541–52. DOI: 10.1016/j.beem.2010.05.007.
140. Edlow A.G., Glass R.M., Smith C.J. et al. Placental Macrophages: A Window Into Fetal Microglial Function in Maternal Obesity. *Int. J. Dev. Neurosci.* 2019; 77, 60–8. DOI: 10.1016/j.ijdevneu.2018.11.004.

141. Хавкин А.И., Айрумов В.А., Шведкина Н.О., Новикова В.П. Биологическая роль и клиническое значение нейрпептидов в педиатрии: пептид YY и грелин. Вопросы практической педиатрии. 2020; 15(5): 87–92. DOI: 10.20953/1817-7646-2020-5-87-92.
142. Symonds M.E., Sebert S.P., Hyatt M.A., Budge H. Nutritional programming of the metabolic syndrome. *Nat Rev Endocrinol.* 2009; 5(11): 604–10. DOI: 10.1038/nrendo.2009.195.
143. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. *Cell.* 2001; 104(4): 531–43. DOI: 10.1016/S0092-8674(01)00240-9.
144. Hassink S.G., Sheslow D.V., de Lancey E. et al. Serum leptin in children with obesity: relationship to gender and development. *Pediatrics.* 1996; 98(2 Pt 1): 201–3.
145. Spiegelman B.M., Flier J.S. Obesity and the regulation of energy balance. *Cell.* 2001; 104(4): 531–43. DOI: 10.1016/S0092-8674(01)00240-9.
146. Morton G.J., Schwartz M.W. Leptin and the central nervous system control of glucose metabolism. *Physiol Rev.* 2011; 91(2): 389–411. DOI: 10.1152/physrev.00007.2010.
147. Hoggard N., Hunter L., Duncan J.S. et al. Leptin and leptin receptor mRNA and protein expression in the murine fetus and placenta. *Proc Natl Acad Sci U S A.* 1997; 94(20): 11073–8. DOI: 10.1073/pnas.94.20.11073.
148. Shekhawat P.S., Garland J.S., Shivpuri C. et al. Neonatal cord blood leptin: its relationship to birth weight, body mass index, maternal diabetes, and steroids. *Pediatr Res.* 1998; 43(3): 338–43. DOI: 10.1203/00006450-199803000-00005.
149. Kamimae-Lanning A.N., Krasnow S.M., Goloviznina N.A. et al. Maternal high-fat diet and obesity compromise fetal hematopoiesis. *Mol. Metab.* 2015; 4(1): 25–38. DOI: 10.1016/j.molmet.2014.11.001.
150. Boeke C.E., Mantzoros C.S., Hughes M.D. et al. Differential associations of leptin with adiposity across early childhood. *Obesity (Silver Spring).* 2013; 21(7): 1430–7. DOI: 10.1002/oby.20314.
151. Mantzoros C.S., Rifas-Shiman S.L., Williams C.J. et al. Cord blood leptin and adiponectin as predictors of adiposity in children at 3 years of age: a prospective cohort study. *Pediatrics.* 2009; 123(2): 682–9. DOI: 10.1542/peds.2008-0343.
152. Ong K.K., Ahmed M.L., Sherriff A. et al. Cord blood leptin is associated with size at birth and predicts infancy weight gain in humans. *ALSPAC Study Team. Avon Longitudinal Study of Pregnancy and Childhood. J Clin Endocrinol Metab.* 1999; 84(3): 1145–8. DOI: 10.1210/jcem.84.3.5657.
153. Zuo H.J., Xie Z.M., Zhang W.W. et al. Gut bacteria alteration in obese people and its relationship with gene polymorphism. *World J. Gastroenterol.* 2011; 17: 1076–81. DOI: 10.3748/wjg.v17.i8.1076.
154. Castro-Rodriguez J.A., Forno E., Casanello P. et al. Leptin in Cord Blood Associates with Asthma Risk at Age 3 in the Offspring of Women with Gestational Obesity. *Ann. Am. Thorac. Soc.* 2020; 17 (12): 1583–9. DOI: 10.1513/AnnalsATS.202001-080OC.
155. Chang G-Q., Gaysinskaya V., Karatayev O. et al. Maternal High-Fat Diet and Fetal Programming: Increased Proliferation of Hypothalamic Peptide-Producing Neurons That Increase Risk for Overeating and Obesity. *J Neurosci.* 2008; 28: 12107–19.
156. Nguyen L.T., Saad S., Tan Y. et al. Maternal high-fat diet induces metabolic stress response disorders in offspring hypothalamus. *J Mol Endocrinol.* 2017; 59: 81–92.
157. Kirk S.L., Samuelsson A-M., Argenton M. et al. Maternal Obesity Induced by Diet in Rats Permanently Influences Central Processes Regulating Food Intake in Offspring. *PLOS ONE.* 2009; 4: e5870.
158. Glavas M.M., Kirigiti M.A., Xiao X.Q. et al. Early overnutrition results in early-onset arcuate leptin resistance and increased sensitivity to high-fat diet. *Endocrinology.* 2010; 151: 1598–1610.
159. Long N.M., Ford S.P., Nathanielsz P.W. Maternal obesity eliminates the neonatal lamb plasma leptin peak. *J Physiol.* 2011; 589: 1455–62.
160. Macumber I., Schwartz S., Leca N. Maternal obesity is associated with congenital anomalies of the kidney and urinary tract in offspring. *Pediatr Nephrol.* 2017; 32: 635–42.
161. Hsu C.W., Yamamoto K.T., Henry R.K. et al. Prenatal risk factors for childhood CKD. *J Am Soc Nephrol JASN.* 2014; 25: 2105–11.
162. Lee Y.Q., Lumbers E.R., Oldmeadow C. et al. The relationship between maternal adiposity during pregnancy and fetal kidney development and kidney function in infants: the Gomeri gaaynggal study. *Physiol Rep.* 2019; 7: e14227.
163. Luyckx V.A., Brenner B.M. The clinical importance of nephron mass. *J Am Soc Nephrol JASN.* 2010; 21: 898–910.
164. Brenner B.M., Chertow G.M. Congenital oligonephropathy and the etiology of adult hypertension and progressive renal injury. *Am J Kidney Dis Off J Natl Kidney Found.* 1994; 23: 171–5.
165. Zhou P., Guan H., Guo Y. et al. Maternal High-Fat Diet Programs Renal Peroxisomes and Activates NLRP3 Inflammasome-Mediated Pyroptosis in the Rat Fetus. *J Inflamm Res.* 2021; 14: 5095–5110.
166. Shamseldeen A.M., Ali Eshra M., Ahmed Rashed L. et al. Omega-3 attenuates high fat diet-induced kidney injury of female rats and renal program-

- ming of their offsprings. *Arch Physiol Biochem.* 2019; 125: 367–77.
167. Glastras S.J., Chen H., McGrath R.T. et al. Effect of GLP-1 Receptor Activation on Offspring Kidney Health in a Rat Model of Maternal Obesity. *Sci Rep.* 2016; 6: 23525.
168. Glastras S.J., Tsang M. Teh R. et al. Maternal Obesity Promotes Diabetic Nephropathy in Rodent Offspring. *Sci Rep.* 2016; 6: 27769.
169. Yamada-Obara N., Yamagishi S., Taguchi K. et al. Maternal exposure to high-fat and high-fructose diet evokes hypoadiponectinemia and kidney injury in rat offspring. *Clin Exp Nephrol.* 2016; 20: 853–61.
170. Ley R.E., Bäckhed F., Turnbaugh P. et al. Obesity alters gut microbial ecology. *Proc. Natl. Acad. Sci. USA.* 2005; 102: 11070–5. DOI: 10.1073/pnas.0504978102.
171. Nguyen L.T., Mak C.H., Chen H. et al. SIRT1 Attenuates Kidney Disorders in Male Offspring Due to Maternal High-Fat Diet. *Nutrients.* 2019; 11: 146.
172. Jackson C.M., Alexander B.T., Roach L. et al. Exposure to maternal overnutrition and a high-fat diet during early postnatal development increases susceptibility to renal and metabolic injury later in life. *Am J Physiol-Ren Physiol.* 2012; 302: F774–F783.
173. Flynn E.R., Alexander B.T., Lee J. et al. High-fat/fructose feeding during prenatal and postnatal development in female rats increases susceptibility to renal and metabolic injury later in life. *Am J Physiol-Regul Integr Comp Physiol.* 2013; 304: R278–R285.
174. Preveden T., Scarpellini E., Milić N. et al. Gut microbiota changes and chronic hepatitis C virus infection. *Expert Rev. Gastroenterol. Hepatol.* 2017; 11: 813–9. DOI: 10.1080/17474124.2017.1343663.
175. Moore W.E., Holdeman L.V. Human fecal flora: The normal flora of 20 Japanese-Hawaiians. *Appl. Microbiol.* 1974; 27: 961–79.
176. Gill S.R., Pop M., Deboy R.T. et al. Metagenomic analysis of the human distal gut microbiome. *Science.* 2006; 312: 1355–9. DOI: 10.1126/science.1124234.
177. Armougom F., Henry M., Viallettes B. et al. Monitoring bacterial community of human gut microbiota reveals an increase in *Lactobacillus* in obese patients and *Methanogens* in anorexic patients. *PLoS ONE.* 2009; 4: e7125. DOI: 10.1371/journal.pone.0007125.
178. Nash A.K., Auchtung T.A., Wong M.C. et al. The gut mycobiome of the Human Microbiome Project healthy cohort. *Microbiome.* 2017; 5: 153. DOI: 10.1186/s40168-017-0373-4.
179. Turnbaugh P.J., Gordon J.I. The core gut microbiome, energy balance and obesity. *J. Physiol.* 2009; 587: 4153–8. DOI: 10.1113/jphysiol.2009.174136.
180. De Faria Ghetti F., Oliveira D.G., de Oliveira J.M. et al. Influence of gut microbiota on the development and progression of nonalcoholic steatohepatitis. *Eur. J. Nutr.* 2018; 57: 861–76. DOI: 10.1007/s00394-017-1524-x.
181. Ottman N., Smidt H., de Vos W.M., Belzer C. The function of our microbiota: Who is out there and what do they do? *Front. Cell. Infect. Microbiol.* 2012; 2: 104. DOI: 10.3389/fcimb.2012.00104.
182. Sittipo P., Lobionda S., Lee Y.K., Maynard C.L. Intestinal microbiota and the immune system in metabolic diseases. *J. Microbiol.* 2018; 56: 154–62. DOI: 10.1007/s12275-018-7548-y.
183. Koleva P.T., Kim J.S., Scott J.A. and Kozyrskyj A.L. Microbial programming of health and disease starts during fetal life. *Birth Defects Res. C Embryo Today.* 2015; 105: 265–77. DOI: 10.1002/bdrc.21117.
184. Escherich T. The intestinal bacteria of the neonate and breast-fed infant. 1885. *Rev. Infect. Dis.* 1989; 11: 352–6. DOI: 10.1093/clinids/11.2.352.
185. Küstner O. Beitrag zur lehre von der puerperalen infection der neugeborenen. *Archiv. für Gynäkologie.* 1877; 11: 256–63. DOI: 10.1007/BF01845161.
186. Perez-Munoz M.E., Arrieta M.C., Ramer-Tait A.E. and Walter J. A critical assessment of the “sterile womb” and “in utero colonization” hypotheses: Implications for research on the pioneer infant microbiome. *Microbiome.* 2017; 5: 48. DOI: 10.1186/s40168-017-0268-4.
187. Collado M.C., Rautava S., Aakko J. et al. Human gut colonisation may be initiated in utero by distinct microbial communities in the placenta and amniotic fluid. *Sci. Rep.* 2016; 6: 23129. DOI: 10.1038/srep23129.
188. Wassenaar T.M. and Panigrahi P. Is a foetus developing in a sterile environment? *Lett. Appl. Microbiol.* 2014; 59: 572–9. DOI: 10.1111/lam.12334.
189. Jimenez E., Marin M.L., Martin R. et al. Is meconium from healthy newborns actually sterile? *Res. Microbiol.* 2008; 159: 187–93. DOI: 10.1016/j.resmic.2007.12.007.
190. Hu J., Nomura Y., Bashir A. et al. Diversified microbiota of meconium is affected by maternal diabetes status. *PLoS One.* 2013; 8: e78257. DOI: 10.1371/journal.pone.0078257.
191. Zheng J., Xiao X.H., Zhang Q. et al. Correlation of placental microbiota with fetal macrosomia and clinical characteristics in mothers and newborns. *Oncotarget.* 2017; 8: 82314–25.
192. Aagaard K., Ma J., Antony K.M. et al. The placenta harbors a unique microbiome. *Sci. Transl. Med.* 2014; 6: 237ra265.
193. Gomez-Arango L.F., Barrett H.L., McIntyre H.D. et al. Contributions of the maternal oral and gut microbiome to placental microbial colonization in

- overweight and obese pregnant women. *Sci. Rep.* 2017; 7: 2860. DOI: 10.1038/s41598-017-03066-4.
194. Mackie R.I., Sghir A., Gaskins H.R. Developmental microbial ecology of the neonatal gastrointestinal tract. In *American Journal of Clinical Nutrition*. 1999; 69(5): 1035S-1045S. June 1999 with 629 Reads.
 195. Hesla H.M., Stenius F., Jäderlund L. et al. Impact of lifestyle on the gut microbiota of healthy infants and their mothers—the ALADDIN birth cohort. *Microbiol Ecol.* 2014; 90 (3): 791–801.
 196. Юдина Ю.В., Аминова А.И., Продеус А.П. и др. Особенности микробиоты кишечника у детей в возрасте 1–5 лет с атопическим дерматитом. *Вопросы детской диетологии*. 2021; 19(2): 5–13.
 197. Jakobsson H.E., Abrahamsson T.R., Jenmalm M.C. et al. Decreased gut microbiota diversity, delayed bacteroidetes colonisation and reduced Th1 responses in infants delivered by caesarean section. *Gut.* 2014; 63(4): 559–66.
 198. MacIntyre D.A., Chandiramani M., Lee Y.S. et al. The vaginal microbiome during pregnancy and the postpartum period in a European population. *Sci Rep.* 2015; 5: 8988.
 199. Jost T., Lacroix C., Braegger C., Chassard C. Assessment of bacterial diversity in breast milk using culture-dependent and culture-independent approaches. *Br J Nutr.* 2013; 110(7): 1253–62.
 200. Soto A., Martín V., Jiménez E. et al. Lactobacilli and bifidobacteria in human breast milk: influence of antibiotherapy and other host and clinical factors. *J Pediatr Gastroenterol Nutr.* 2014; 59(1): 78–88.
 201. Zivkovic A.M., German J.B., Lebrilla C.B., Mills D.A. Human milk glycobiome and its impact on the infant gastrointestinal microbiota. *Proc Natl Acad Sci USA.* 2011; 108(1): 4653–61.
 202. Garrido D., Ruiz-Moyano S., Mills D.A. Release and utilization of N-acetyl-D-glucosamine from human milk oligosaccharides by *Bifidobacterium longum* subsp. *Infantis* Anaerobe. 2012; 18(4): 430–35.
 203. Penders J., Thijs C., Vink C. et al. Factors influencing the composition of the intestinal microbiota in early infancy. *Pediatrics.* 2006; 118(2): 511–21.
 204. Rutayisire E., Huang K., Liu Y., Tao F. The mode of delivery affects the diversity and colonization pattern of the gut microbiota during the first year of infants' life: a systematic review. *BMC Gastroenterology.* 2016; 16: 86.
 205. Panda S., El khader I., Casellas F. et al. Short-term effect of antibiotics on human gut microbiota. *PLoS One.* 2014; 9(4): e95476.
 206. Sonnenburg J.L., Sonnenburg E.D. Vulnerability of the industrialized microbiota. *Science.* 2019; 366: eaaw9255.
 207. Stark C.M., Susi A., Emerick J., Nylund C.M. Antibiotic and acid-suppression medications during early childhood are associated with obesity. *Gut.* 2019; 68: 62–9.
 208. Kronman M.P., Zaoutis T.E., Haynes K. et al. Antibiotic exposure and iBD development among children: A population-based cohort study. *Pediatrics* 2012; 130: e794–e803.
 209. Langdon A., Crook N., Dantas G. The effects of antibiotics on the microbiome through out development and alternative approaches for therapeutic modulation. *Genome Med.* 2016; 8(1): 39.
 210. Turnbaugh P.J., Ley R.E., Mahowald M.A. et al. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature.* 2006; 444: 1027–31. DOI: 10.1038/nature05414.
 211. Schwiertz A., Taras D., Schäfer K. et al. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity.* 2010; 18: 190–5. DOI: 10.1038/oby.2009.167.
 212. Turnbaugh P.J., Quince C., Faith J.J. et al. Organismal, genetic, and transcriptional variation in the deeply sequenced gut microbiomes of identical twins. *Proc. Natl. Acad. Sci. USA.* 2010; 107: 7503–8. DOI: 10.1073/pnas.1002355107.
 213. Waldram A., Holmes E., Wang Y. et al. Top-down systems biology modeling of host metabolite-microbiome associations in obese rodents. *J. Proteome Res.* 2009; 8: 2361–75. DOI: 10.1021/pr8009885.
 214. Zhang H., DiBaise J.K., Zuccolo A. et al. Human gut microbiota in obesity and after gastric bypass. *Proc. Natl. Acad. Sci. USA.* 2009; 106: 2365–70. DOI: 10.1073/pnas.0812600106.
 215. Cuevas-Sierra A., Ramos-Lopez O., Riezu-Boj J.I. et al. Diet, Gut Microbiota, and Obesity: Links with Host Genetics and Epigenetics and Potential Applications. *Adv. Nutr.* 2019; 10: S17–S30. DOI: 10.1093/advances/nmy078.
 216. Hiippala K., Jouhten H., Ronkainen A. et al. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients.* 2018; 10: 988. DOI: 10.3390/nu10080988.
 217. Cani P.D., Amar J., Iglesias M.A. et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes.* 2007; 56: 1761–72. DOI: 10.2337/db06-1491.
 218. Xu P., Li M., Zhang J., Zhang T. Correlation of intestinal microbiota with overweight and obesity in Kazakh school children. *BMC Microbiol.* 2012; 12: 283. DOI: 10.1186/1471-2180-12-283.
 219. Munukka E., Wiklund P., Pekkala S. et al. Women with and without metabolic disorder differ in their gut microbiota composition. *Obesity.* 2012; 20: 1082–7. DOI: 10.1038/oby.2012.8.

220. Kravtsova K., Prokopieva N.E., Petrenko Yu.V. et al. Gut microbiota of children born to obese mothers. *World of Microbiome*, Vienna, 28–30 апреля 2022 года. Vienna: Kenes group. 2022.
221. Gagliardi A., Totino V., Cacciotti F. et al. Rebuilding the Gut Microbiota Ecosystem. *Int. J. Environ. Res. Public Health*. 2018; 15: 1679. DOI: 10.3390/ijerph15081679.
222. Zhao Y., He X., Shi X. et al. Association between serum amyloid A and obesity: A meta-analysis and systematic review. *Inflamm. Res*. 2010; 59: 323–34. DOI: 10.1007/s00011-010-0163-y.
223. Payne A.N., Chassard C., Zimmermann M. et al. The metabolic activity of gut microbiota in obese children is increased compared with normal-weight children and exhibits more exhaustive substrate utilization. *Nutr. Diabetes*. 2011; 1: e12. DOI: 10.1038/nutd.2011.8.
224. Alex S., Lichtenstein L., Dijk W. et al. ANGPTL4 is produced by entero-endocrine cells in the human intestinal tract. *Histochem. Cell Biol*. 2014; 141: 383–91. DOI: 10.1007/s00418-013-1157-y.
225. Sanmiguel C., Gupta A., Mayer E.A. Gut Microbiome and Obesity: A Plausible Explanation for Obesity. *Curr. Obes. Rep*. 2015; 4: 250–61. DOI: 10.1007/s13679-015-0152-0.
226. Stephens R.W., Arhire L., Covasa M. Gut Microbiota: From Microorganisms to Metabolic Organ Influencing Obesity. *Obesity*. 2018; 26: 801–9. DOI: 10.1002/oby.22179.
227. Bäckhed F., Ding H., Wang T. et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. USA*. 2004; 101: 15718–23. DOI: 10.1073/pnas.0407076101.
228. Scarpellini E., Cazzato A., Lauritano C. et al. Probiotics: Which and when? *Dig. Dis*. 2008; 26: 175–82. DOI: 10.1159/000116776.
229. Смирнова Н.Н., Новикова В.П., Куприенко Н.Б. и др. Влияние микробиома репродуктивного тракта женщины на внутриутробное и постнатальное развитие ребенка. *Вопросы гинекологии, акушерства и перинатологии*. 2022; 21(6): 107–13. DOI: 10.20953/1726-1678-2022-6-107-112. EDN LCUFFM.
230. Duncan S.H., Belenguer A., Holtrop G. Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate-producing bacteria in feces. *Appl. Environ. Microbiol*. 2007; 73: 1073–8. DOI: 10.1128/AEM.02340-06.
231. Murphy E.F., Cotter P.D., Healy S. et al. Composition and energy harvesting capacity of the gut microbiota: Relationship to diet, obesity and time in mouse models. *Gut*. 2010; 59: 1635–42. DOI: 10.1136/gut.2010.215665.
232. Li J.V., Ashrafi H., Bueter M. et al. Metabolic surgery profoundly influences gut microbial-host metabolic cross-talk. *Gut*. 2011; 60: 1214–23. DOI: 10.1136/gut.2010.234708.
233. Liou A.P., Paziuk M., Luevano J.M., Jr. et al. Conserved shifts in the gut microbiota due to gastric bypass reduce host weight and adiposity. *Sci. Transl. Med*. 2013; 5: 178ra41. DOI: 10.1126/scitranslmed.3005687.
234. Hiippala K., Jouhten H., Ronkainen A. et al. The Potential of Gut Commensals in Reinforcing Intestinal Barrier Function and Alleviating Inflammation. *Nutrients*. 2018; 10: 988. DOI: 10.3390/nu10080988.
235. Cani P.D., Amar J., Iglesias M.A. et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007; 56: 1761–72. DOI: 10.2337/db06-1491.
236. Lee S., Sung J., Lee J., Ko G. Comparison of the gut microbiotas of healthy adult twins living in South Korea and the United States. *Appl. Environ. Microbiol*. 2011; 77: 7433–7. DOI: 10.1128/AEM.05490-11.
237. Luoto R., Kalliomäki M., Laitinen K. et al. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J. Pediatr. Gastroenterol. Nutr*. 2011; 52: 90–5. DOI: 10.1097/MPG.0b013e-3181f3457f.
238. Payne A.N., Chassard C., Banz Y., Lacroix C. The composition and metabolic activity of child gut microbiota demonstrate differential adaptation to varied nutrient loads in an in vitro model of colonic fermentation. *FEMS Microbiol. Ecol*. 2012; 80: 608–23. DOI: 10.1111/j.1574-6941.2012.01330.x.
239. Gohir W., Whelan F.J., Surette M.G. et al. Pregnancy-related changes in the maternal gut microbiota are dependent upon the mother's periconceptional diet. *Gut Microbes*. 2015; 6: 310–20. DOI: 10.1080/19490976.2015.1086056.
240. Collado M.C., Isolauri E., Laitinen K. and Salminen S. Distinct composition of gut microbiota during pregnancy in overweight and normal-weight women. *Am. J. Clin. Nutr*. 2008; 88: 894–9.
241. Qin J., Li R., Raes J., et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature*. 2010; 464: 59–65. DOI: 10.1038/nature08821.
242. Dietert R.R. and Dietert J.M. The microbiome and sustainable healthcare. *Healthcare (Basel)*. 2015; 3: 100–29. DOI: 10.3390/healthcare3010100.
243. Zacarias M.F., Collado M.C., Gómez-Gallego C. et al. Pregestational overweight and obesity are associated with differences in gut microbiota composition and systemic inflammation in the third trimester. *PLoS One*. 2018; 13(7): e0200305. DOI: 10.1371/journal.pone.0200305. PMID: 30005082; PMCID: PMC6044541.

244. Santacruz A., Collado M.C., Garcia-Valdes L. et al. Gut microbiota composition is associated with body weight, weight gain and biochemical parameters in pregnant women. *Br. J. Nutr.* 2010; 104: 83–92. DOI: 10.1017/S0007114510000176.
245. Soderborg T.K., Clark S.E., Mulligan C.E. et al. The gut microbiota in infants of obese mothers increases inflammation and susceptibility. *Nat. Commun.* 2018; 9(1). DOI: 10.1038/s41467-018-06929-0.
246. Ferrer M., Ruiz A., Lanza F. et al. Microbiota from the distal guts of lean and obese adolescents exhibit partial functional redundancy besides clear differences in community structure. *Environ. Microbiol.* 2013; 15: 211–26. DOI: 10.1111/j.1462-2920.2012.02845.x.
247. Bervoets L., Van Hoorenbeeck K., Kortleven I. et al. Differences in gut microbiota composition between obese and lean children: A cross-sectional study. *Gut Pathog.* 2013; 5: 10. DOI: 10.1186/1757-4749-5-10.
248. Luoto R., Kalliomäki M., Laitinen K. et al. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J. Pediatr. Gastroenterol. Nutr.* 2011; 52: 90–5. DOI: 10.1097/MPG.0b013e-3181f3457f.
249. Clarke S.F., Murphy E.F., O'Sullivan O. et al. Targeting the microbiota to address diet-induced obesity: A time dependent challenge. *PLoS One.* 2013; 8: e65790 DOI: 10.1371/journal.pone.0065790.
250. Прокопьева Н.Э., Петренко Ю.В., Иванов Д.О. и др. Формирование полостной микробиоты кишечника у детей первого года жизни, рожденных от матерей с ожирением. Актуальные проблемы абдоминальной патологии у детей: Материалы Юбилейного XXX Конгресса детских гастроэнтерологов России и стран СНГ, Москва, 14–16 марта 2023 года. М.: Медпрактика-М; 2023: 41–3.
251. Martinez I., Lattimer J.M., Hubach K.L. et al. Gut microbiome composition is linked to whole grain-induced immunological improvements. *Isme J.* 2013; 7: 269–80. DOI: 10.1038/ismej.2012.104.
252. Vael C., Verhulst S.L., Nelen V. et al. Intestinal microbiota and body mass index during the first three years of life: An observational study. *Gut Pathog.* 2011; 3: 8. DOI: 10.1186/1757-4749-3-8.
253. Nadal I., Santacruz A., Marcos A. et al. Shifts in clostridia, bacteroides and immunoglobulin-coating fecal bacteria associated with weight loss in obese adolescents. *Int. J. Obes.* 2009; 33: 758–67. DOI: 10.1038/ijo.2008.260.